

Data Integration

Lectures 16 & 17

Lectures Outline

- Goals for Data Integration
- Homogeneous data integration
 - time series data (Filkov et al. 2002)
- Heterogeneous data integration
 - microarray + sequence
 - microarray + protein
 - microarray + location
 - is integration always beneficial?
- Data Integration for Developmental Networks (Davidson et al., 2002)

Integrating data from various experiments should yield better understanding of the data compared to that of individual data sets.

Goals for Data Integration

We integrate data sets with specific goals in mind:

- better gene classification
- better gene clustering
- better regulatory networks

Methods used are the same (modeling):

- SVMs
- Bayesian inference
- Clustering/Classification
- Graph models and algorithms
- Statistical Significance

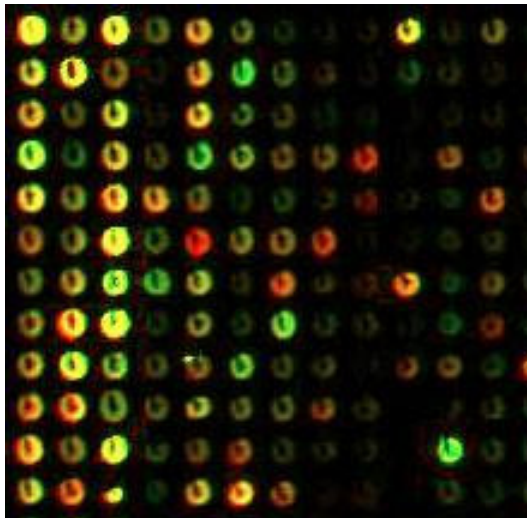
Homogeneous Data

- Expression Data (microarrays)
- Sequence Data
- Location Data (ChIP)
- Protein Expression Data

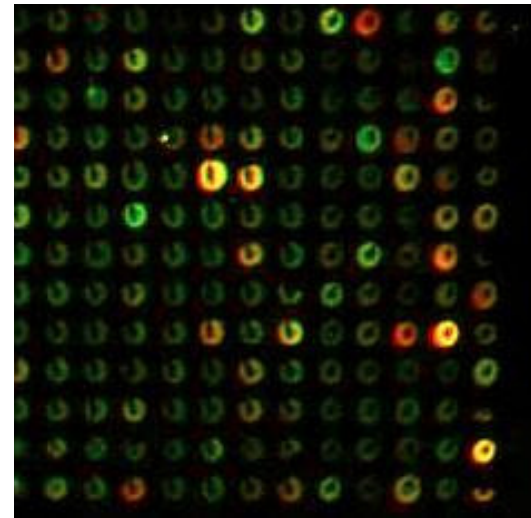
Common platforms for storage, retrieval and comparison across similar data type

Homogeneous Integration

Eg. Microarray expression data is compared across treatments to discover differential gene expression, i.e. genes that behave differently under treatment w.r.t control



treatment



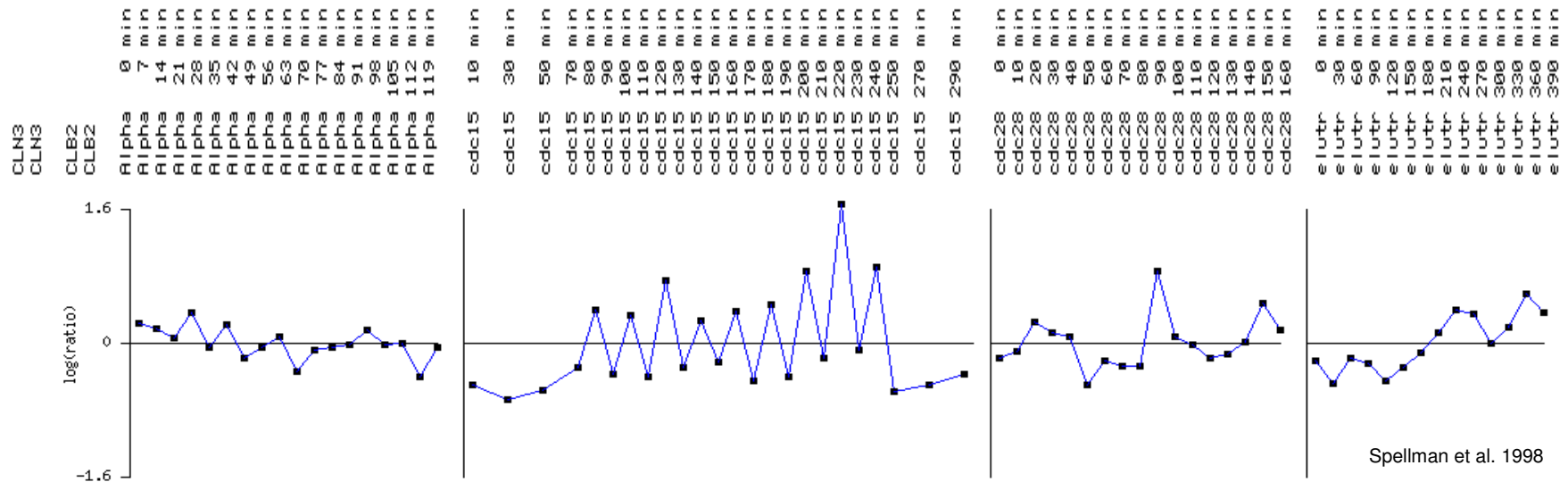
control

Homogeneous Integration: Time-series

(Filkov et al., 2002)

Yeast cells in different experiments are synchronized differently

Plot of ACT1 (YFL039C)

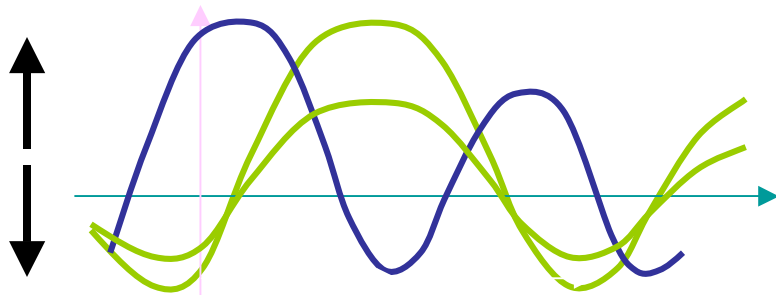
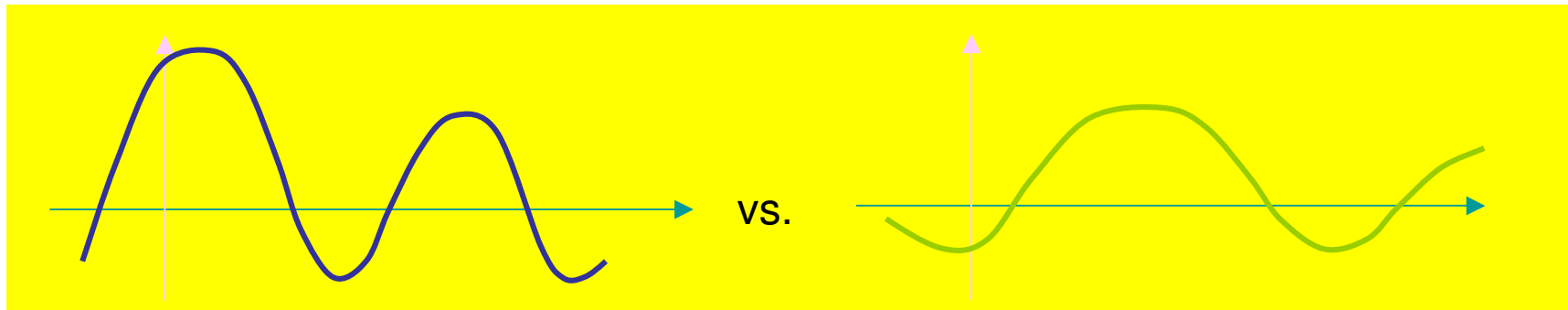


How to integrate time-series data?

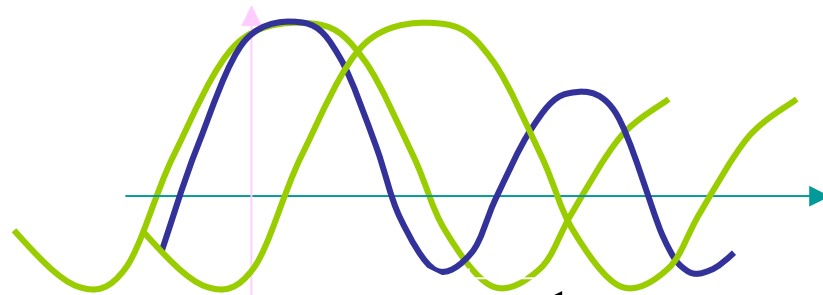
Warp the curves so that they all have the same

- amplitude
- phase and
- period

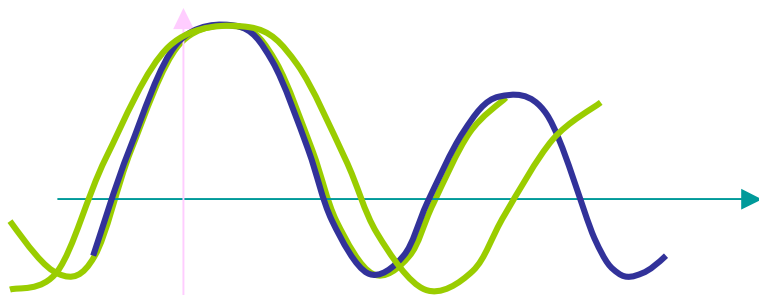
Warping Time-series



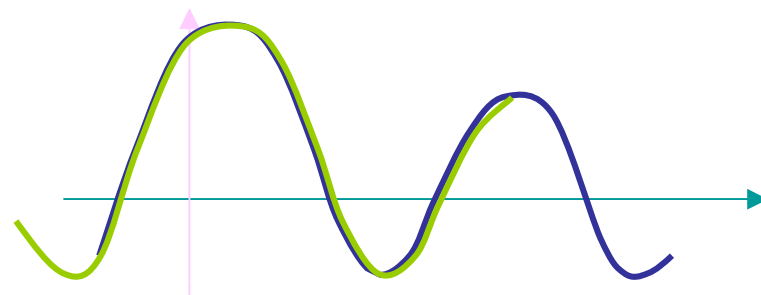
Amplitude



Phase



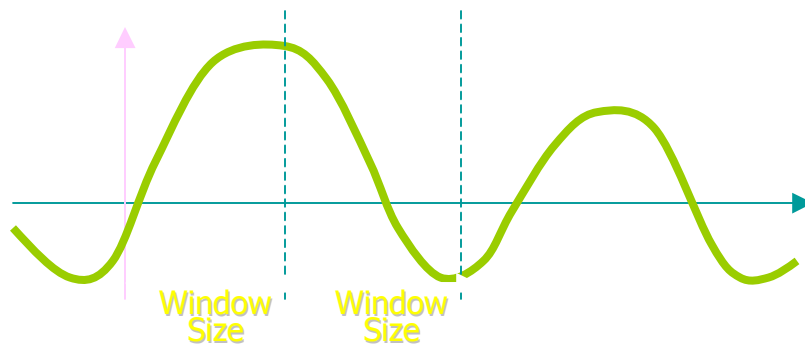
Period



Final

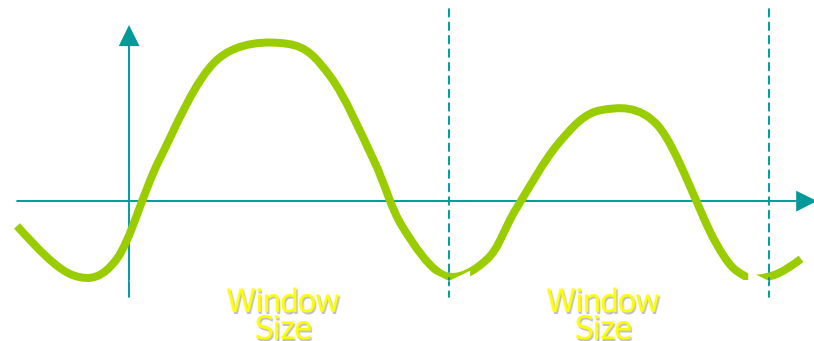
Period and Phase Warp

1. Assume Most Genes are Periodic
2. Perform auto-correlation studies to find period and phase shift
3. Correct for correlation significance in short sequences

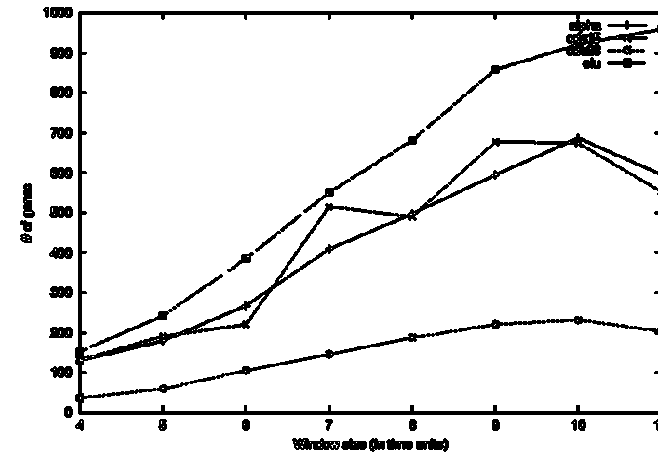
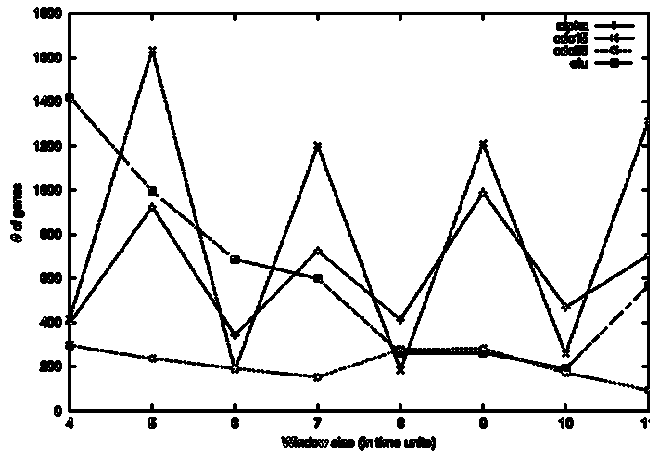


Window length equal to cell cycle length => large correlation

Window length different from cell cycle length => small correlation



After correcting for chance the data sets periods are predicted correctly



Data Set	Period Observed	Period Detected	Dt	# samples	# full orfs
Alpha	$66 \pm 11\text{min}$	$70 \pm 7\text{min}$	7	18	3361
Cdc28 (Cho)	$90 \pm 10\text{min}$	$100 \pm 10\text{min}$	10	17	1188
Cdc15	$70 \pm 10\text{min}$	$90 \pm 10\text{min}$	10/20	24	3453
elu	---	---	30	14	4753

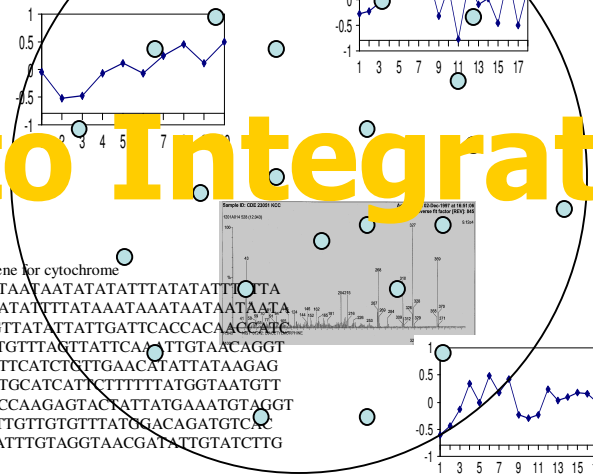
(phase shift determined similarly...)

Heterogeneous Data Integration

- DNA Sequence
- Microarray
- Proteomics

Important to Integrate!

```
>gil12004594|gb|AF217406.1| Saccharomyces cerevisiae uridine nucleosidase (URH1) gene, complete cds
ATGGAATCTGCTGATTTTTTACCTCACGAAACTTATTAACAGATAATTCCTCATCTGCAAGGTTG
GGGAAGGGTTGGACAGCAATAACCAGCGACTGTTTGAAGAAAAGATGACTGTTAGTAAAATACCCATATG
GCTAGATTGTGATCCTGGTCATGATGCCATAGCCATTTTATTAGGCTGTTCCATCCAGCTTCAAT
CTTCTAGGAATCAGCACGTGTTTGGTAACGCACCGCCAGAGAATACTGACTACAACGCCCTTCTCTT
TGAATGCGATGGGCAAAGCACAAAGCGATTCCAGTTTATAAAGGCGCACAGAGACCTTGGAAAAGGGAAC
CTCATTATGCTCCGACATTCATGGTAAATCAGGTTTAGACGGCACTTCTTGTACCTAAGCCAACATTT
GAGGCAAGAAGTGAATAAACGTATATTGAGGCCATTGAAGAGGCGATTCTAGCTAACCAATGGAGAGATA
T CCTTTGTGCTACTGTTCCCTTACCATAGCAACAGTTTTTAGGTGTAACCATACCTAAAAAATC
```



```
>gil13534|embl|V00696.1|MISC16 Yeast (S. cerevisiae) mitochondrial gene for cytochrome
ATATATATAATTATAAATATATATATAATAAGTATTAATTAATAATATATATTTATATATTTTATA
TTAATTAATATATAAAAATATTAGTAATAAATAATATTATTAATAATTTATAAATAAATAATAATAA
TGGCATTAGAAAATCAAATGTGATTTAAGTTTGTAGTAATAGTTATTTATTGATCACCACAACCAATC
ATCAATTAATTATTGATGAAAATATGGGTTTCATTATTAGGTTTATGTTTATTTATTCAAATTTGTAACAGGT
ATTTTTATGGCTATGCATTATTCATCTAATATTGAATTAGCTTTTTTCATCTGTTGAACATATTATAAGAG
ATGTGCATAAATGGTTATATTTAAGATATTTACATGCAAATGGTGCAATCAATTTTTTATGGTAATGTT
TATGCATATGGCTAAAGGTTTATATTATGGTTTATAGATACCAAGAGTACATTTATGAAATGATGGT
GTTATTATTTTCATTTAATGCTACAGCTTTTTAGGTTATTTGTTGTTTATGACAGATGTCATTTG
ATTGAGGTGCACTAGTTATTACTAATTTATTCTCAGCAATTCATTTGTAGGTAACGATATTGTATCTTG
```

Yeast Genes

Why Does It Pay to Integrate?

- Gifford, Computational Functional Genomics, Lecture 18
- “Multiple independent constraints can dramatically increase the significance of otherwise elusive effects”
- Dependent vs. Independent

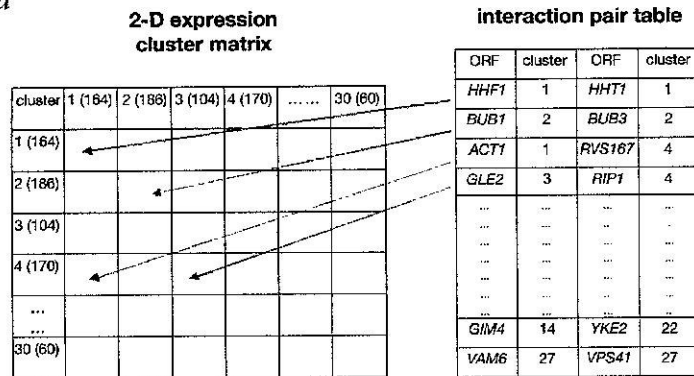
1. Classification

- Simple, intuition based classifications
 - compare to the leukemia classification of Golub et al.,
- Machine Learning classifiers (SVMs)
 - compare to Cristianini et al.

Eg. Gene Expression + Protein Interaction Data

(Ge et al., 2001)

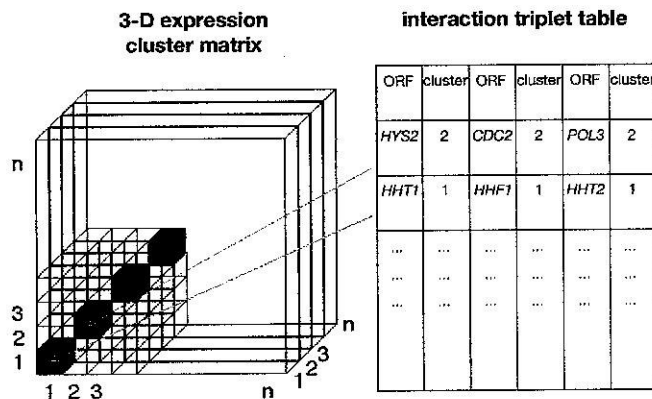
a



Goals

1. To compare the levels of interaction between proteins encoded by co-expressed genes vs. proteins not encoded by co-expressed genes
2. Improved modeling of protein-protein interactions

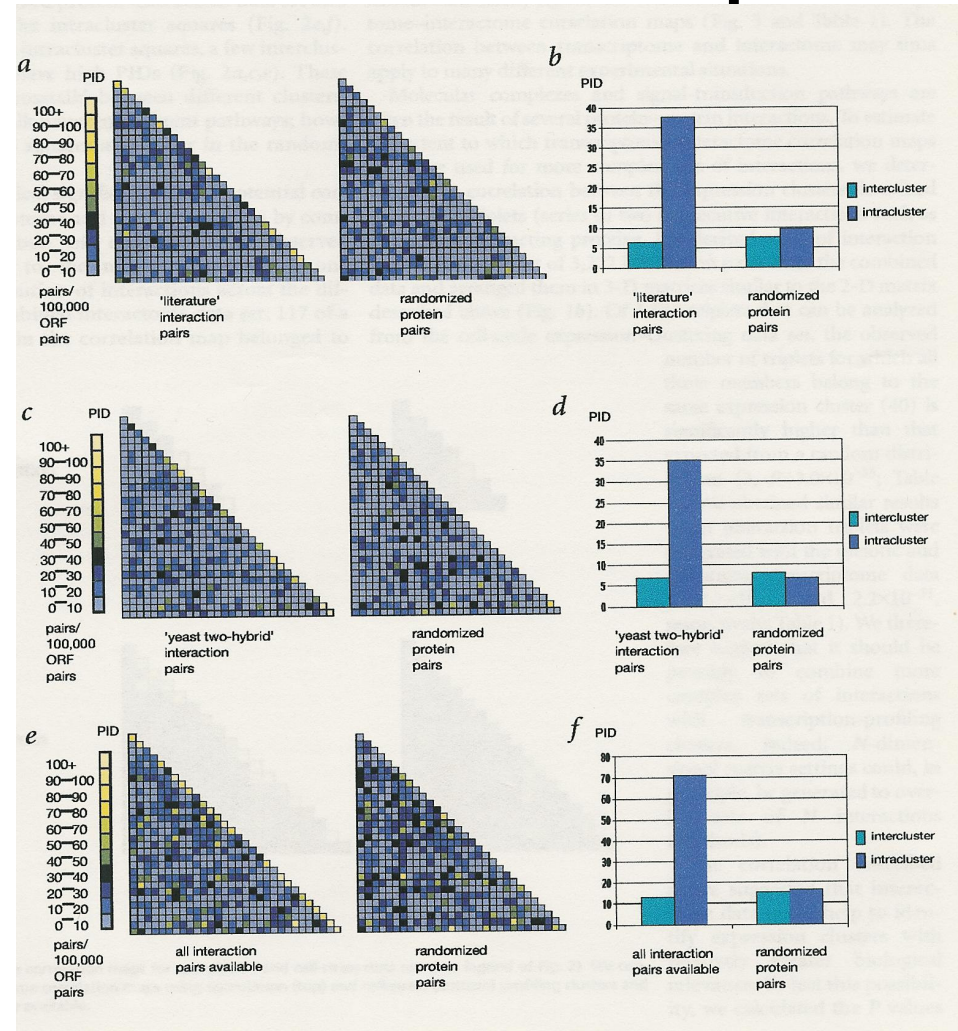
b



Methods

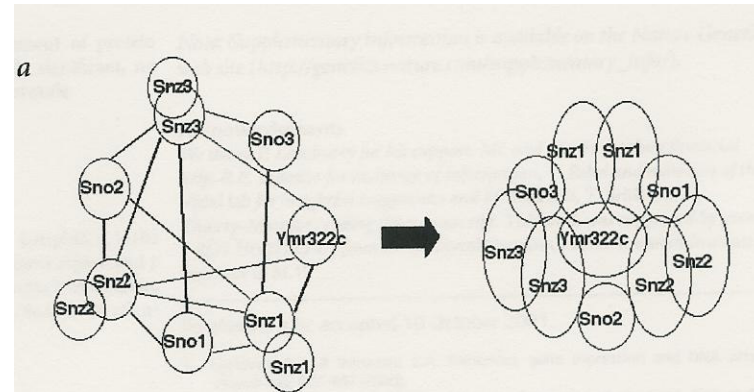
Calculate protein interaction density, and corresponding significance within and between co-expressed clusters of genes

Transcriptome – Interactome Correlation Maps

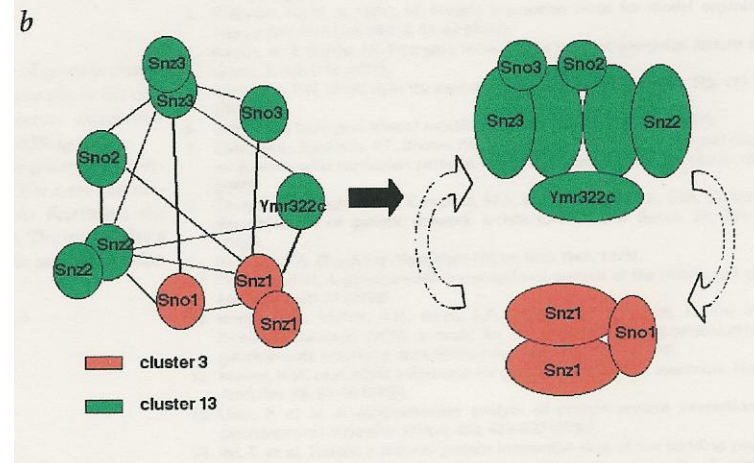


More Knowledge Yields Better Models

a) Protein-protein interaction data



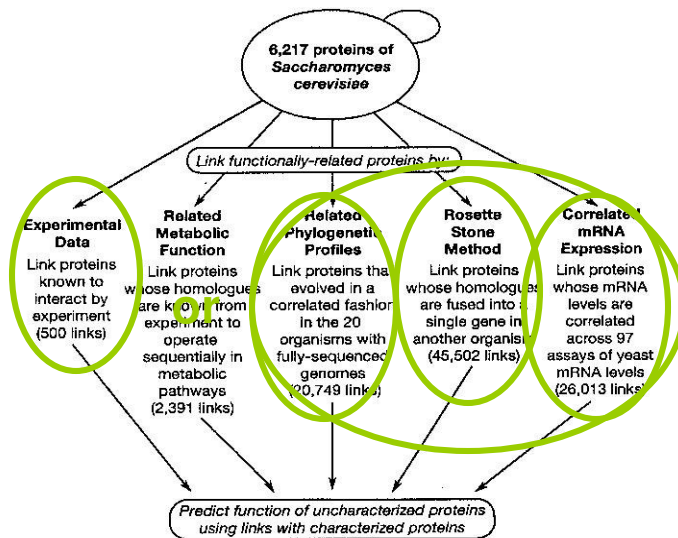
b) Protein Interaction + Gene Expression Data



Stress response proteins

Eg. Protein Function Prediction

(Marcotte et al., 1999)



Combining various strategies to link functionally related proteins. Total: 93750 links

Link confidence:

- highest confidence (4130 links)
- high confidence (19521 links)
- rest

Table 1 Reliability of functional assignments assessed by recovery of known protein function by prediction

	Number of proteins	Number of functional links	False positive rate* (%)	Ability to predict known function† (%)	Ability in random trials‡ (%)	Signal to noise ratio§
Individual prediction techniques						
Experimentall	484	500	6.5	33.2	4.0	8.3
Metabolic pathway neighbours	188	2,391	2.5	20.3	4.5	4.5
Phylogenetic profiles	1,976	20,749	29.5	33.1	7.4	4.5
Rosetta Stone method	1,898	45,502	36.4	26.5	7.7	3.4
Correlated mRNA expression	3,397	26,013	35.8	11.5	6.9	1.7
Combined predictions						
Links made by ≥2 prediction techniques	683	1,249	16.1	55.6	6.9	8.1
Highest confidence links	1,223	4,130	4.8	40.9	5.5	7.4
High confidence links	1,930	19,521	30.6	30.8	7.4	4.2
High and highest confidence links	2,356	23,651	21.8	32.0	6.8	4.7
All links	4,701	93,750	33.1	20.7	7.2	2.9

* The reliability of individual links was calculated as the percentage of pairwise links found between proteins of known function but having no functional categories in common (as tabulated in the MIPS database), ignoring the functional categories 'unclassified' and 'classification not clear cut'. This estimate of false positives assumes complete knowledge of protein function and is therefore an upper limit. By this test, random links achieve a false positive rate of ~47%.

† The predictive power of individual techniques and combinations of techniques was evaluated by automated comparison of annotation keywords. By the methods listed, each protein is linked to one or more neighbour proteins. For characterized proteins ('query' proteins), the mean recovery of known Swiss-Prot keyword annotation by the keyword annotation of linked neighbours was calculated as:

$$\langle \text{keyword recovery} \rangle = \frac{1}{A} \sum_{i=1}^x \frac{n_i}{N} \quad (1)$$

where A is the number of annotated proteins, x is the number of query protein Swiss-Prot keywords, N is the total number of neighbour protein Swiss-Prot keywords, and n_i is the number of times query protein keyword i occurs in the neighbour protein annotation. Because functional annotations typically consist of multiple keywords, both specific and general, even truly related proteins show only a partial keyword overlap (for example, ~35%).

‡ Mean recovery of Swiss-Prot keyword annotation for query proteins of known function by Swiss-Prot keyword annotation of randomly chosen linked neighbours, calculated as in equation (1) for the same number of links as exist for real links (averages of 10 trials).

§ Calculated as ratio of known function recovered by real links to that recovered by random links. Although individual links have only moderate accuracy, combining information from many links significantly enhances prediction of function.

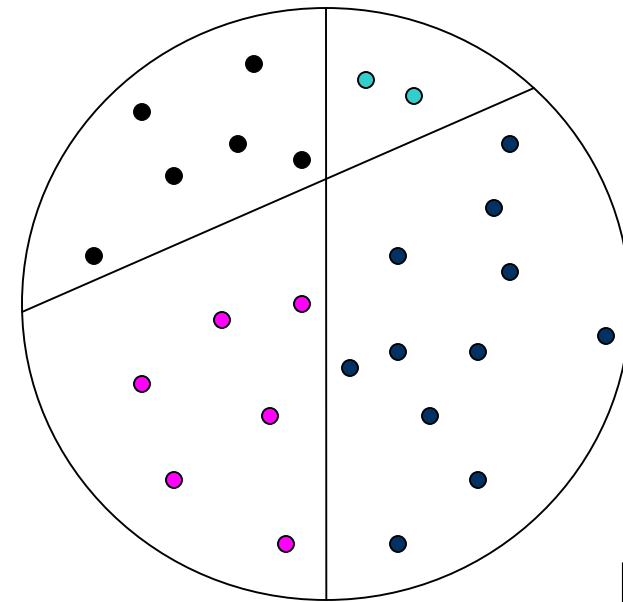
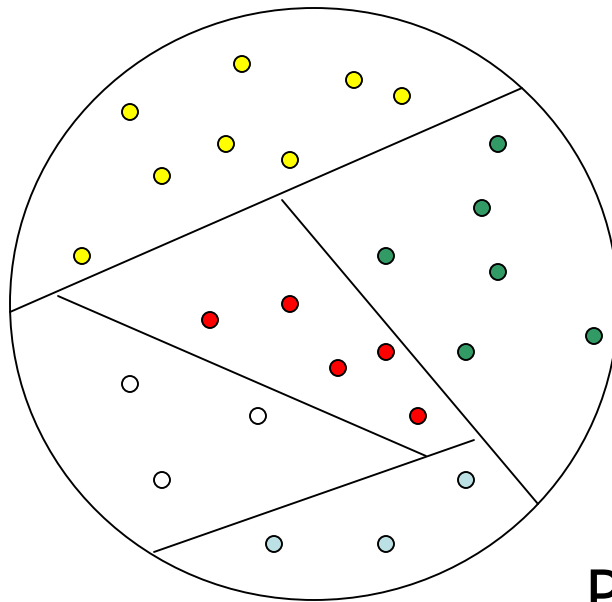
|| Experimentally observed yeast protein-protein interactions contained in the DIP³ and MIPS⁴ databases.

2. Integrating Clusterings

(Filkov and Skiena, 2003)

Data sets are usefully summarized as clusterings

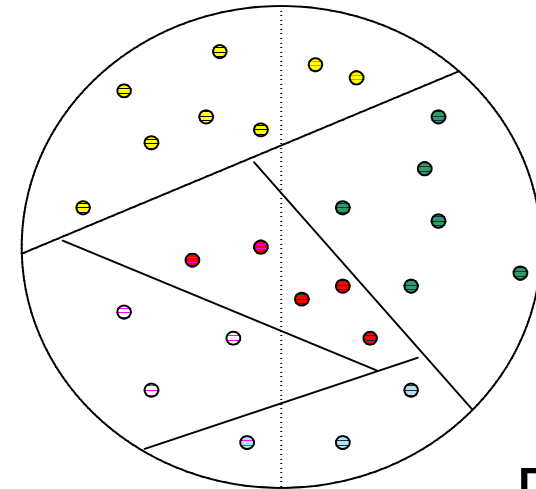
- Functional
- Structural
- Data Driven



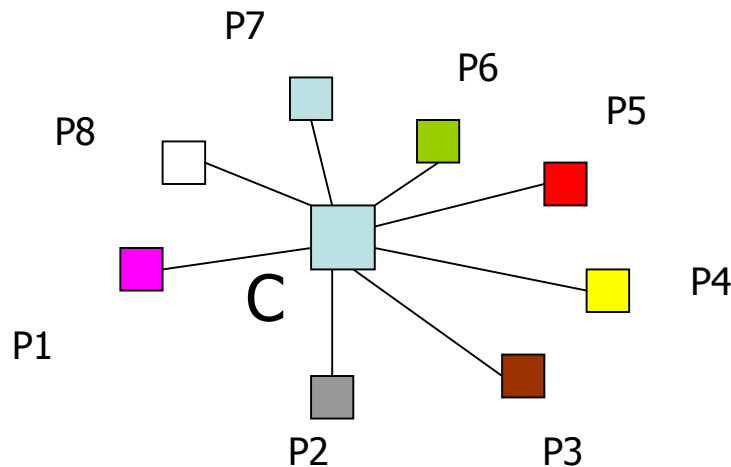
By using multiple clusterings we can learn more, but how?

Overlapping two clusterings is useful, but can we generalize it?

Problem: Find a Consensus Clustering that describes the given clusterings well



P1

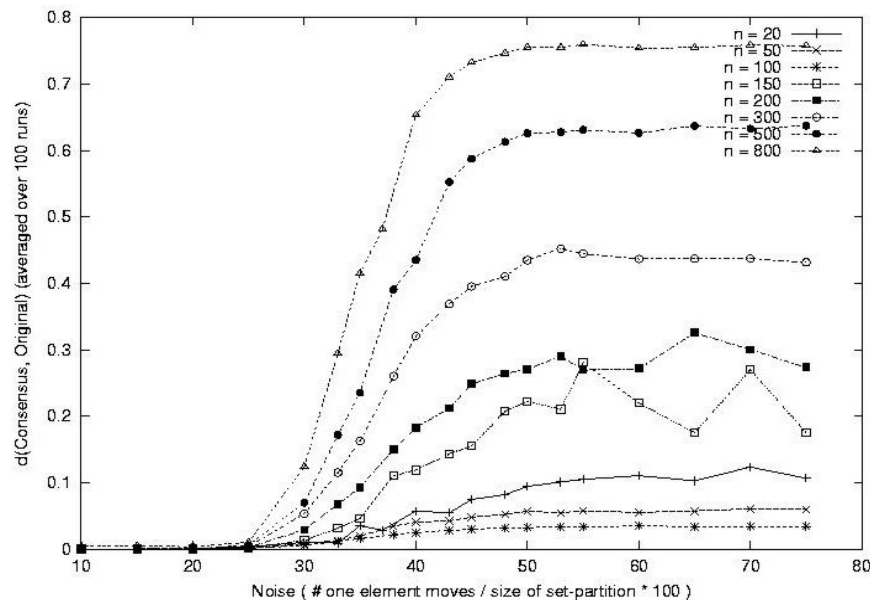


Approach: Integrate the clusterings of data by minimizing the sum of distances between them and a consensus

$$\min S = \sum_i d(P_i, C)$$

Solving the Consensus Problem

- min S consensus is NP-complete even for a very simple distance function (Rand Index)
- Simple Heuristics based on random element move between clusters work well on large data sets
- a measure of benefit of integration



(artificial data)

■ Integrating Spellman's Data

— Alpha, Avg. SoD = 0.1121

— cdc15, Avg. SoD = 0.1042

— elu, Avg. SoD = 0.1073

— Overall, Avg. SoD = 0.107, benefit

■ Spellman + Phylogeny = No benefit

■ Spellman + Yeast Stress = Benefit

(real data)

3. Gene Network Inference

- Data Integration for Link (Graph) Modeling in General
- Probabilistic Setting
- Each data source is an “expert” proposing a model
- Independent experts: easy (Gifford 2002)
 - independent significances, p_1, p_2
 - combined significance, $p=f(p_1, p_2)$

Graph Models

- Dependent experts (Hartemink et al., 2002)
 - Joint probability distributions
 - Bayesian Networks
 - Model scoring
 - Maximizing a Bayesian scoring function
 - simulated annealing optimizer
 - averaging over high-scoring models
 - Location+expression data used as priors

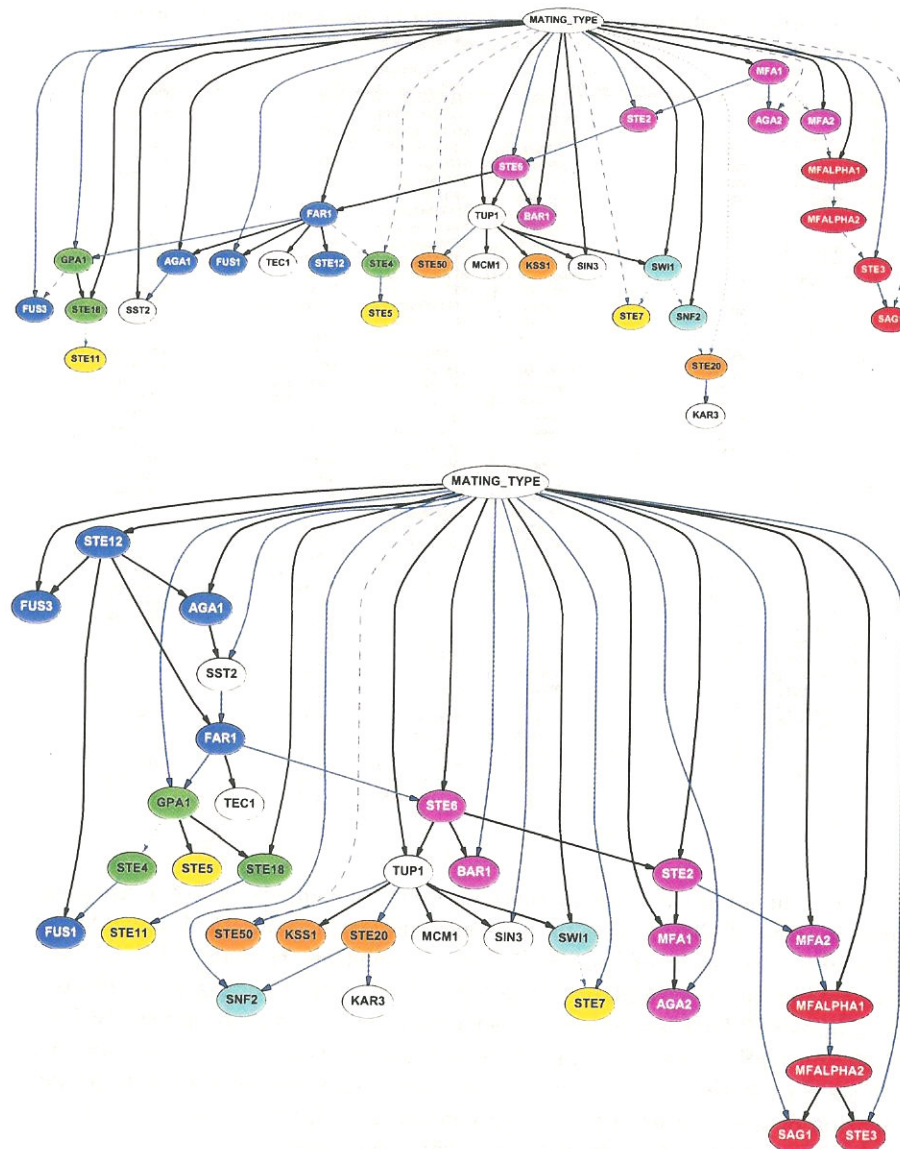
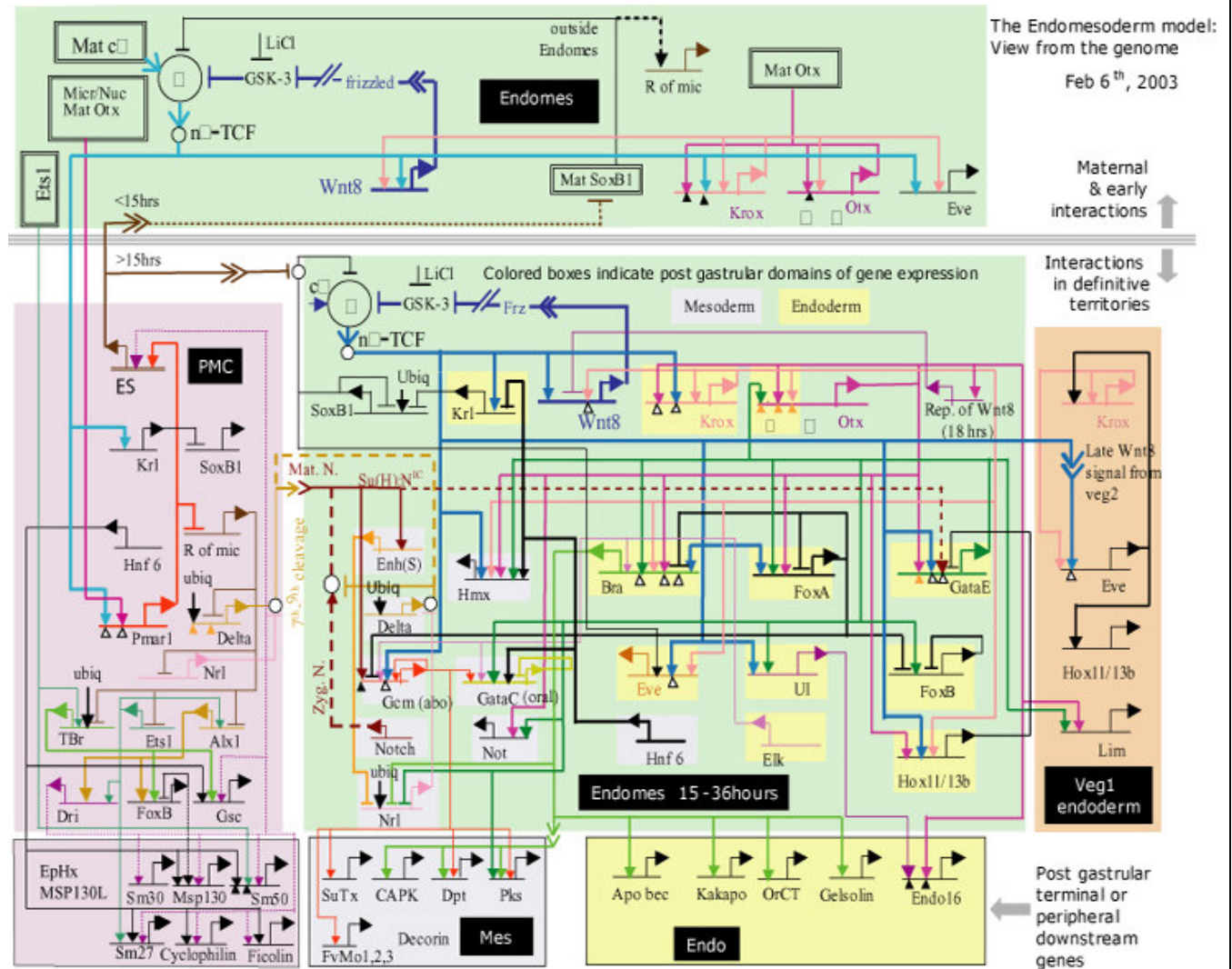


Figure 2. Bayesian network models learned by model averaging over the 500 highest scoring models visited during the unconstrained and constrained simulated annealing search runs, respectively. Edges are included in the figure if and only if their posterior probability exceeds 0.5. Node and edge color descriptions are included in the text.

4. Putting It All Together

Davidson et al., 2002



Bibliography

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