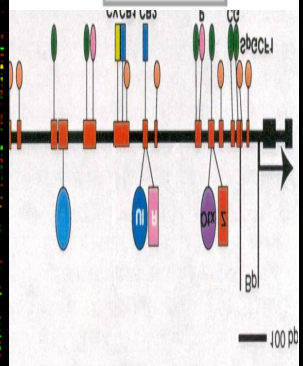
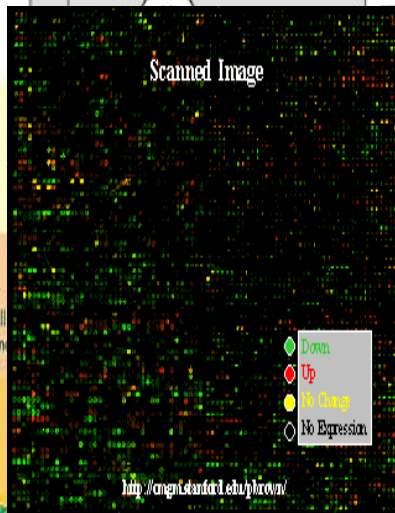
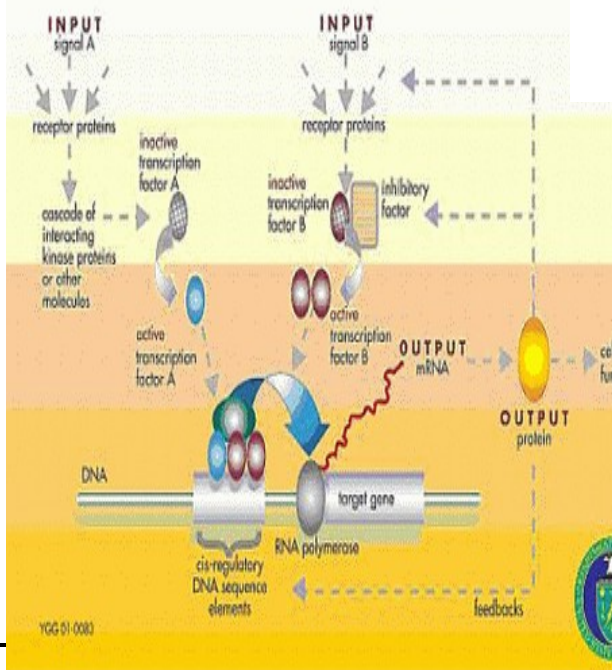
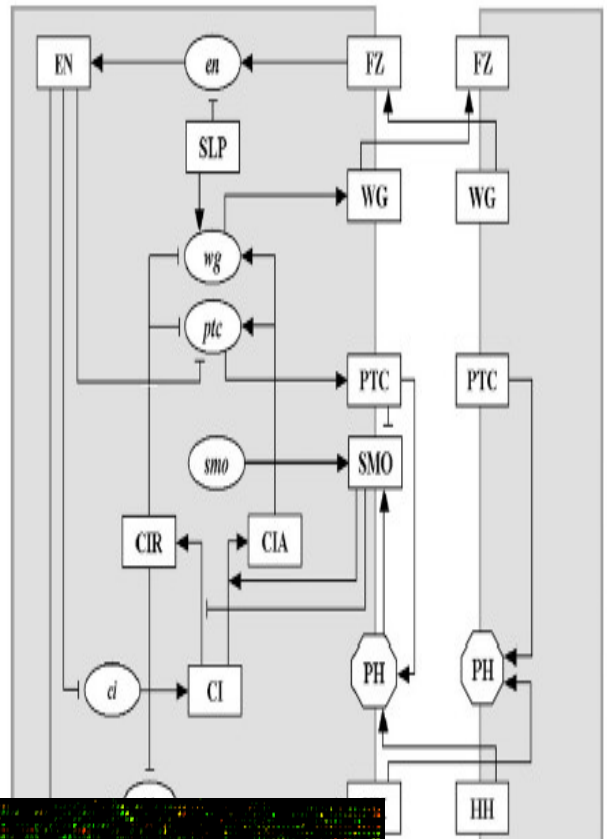
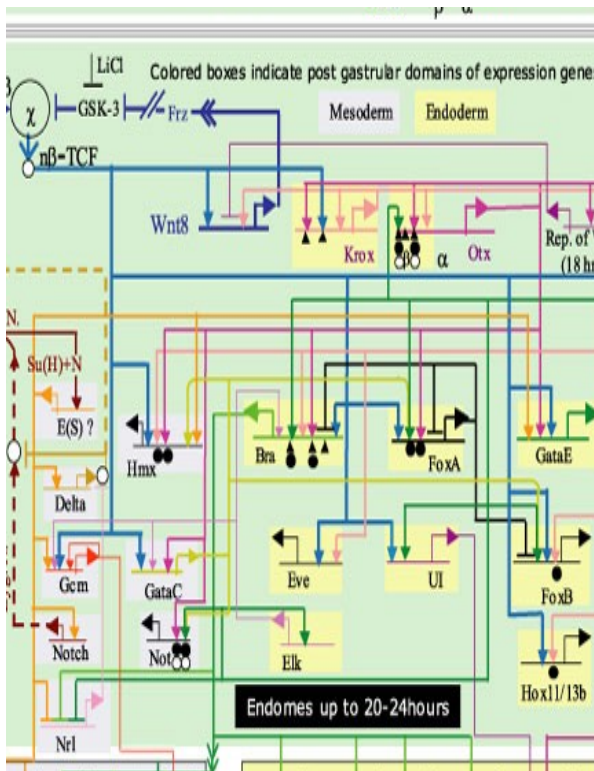


# ECS 234: Gene Regulation Modelling: Continuous Models



# Transcriptional Regulatory Systems

- Cis regulatory elements: DNA sequence (specific sites)
  - promoters;
  - enhancers;
  - silencers;
- Trans regulatory factors: products of regulatory genes
  - generalized
  - specific (Zinc finger, leucine zipper, etc.)

Known properties of real gene regulatory systems:

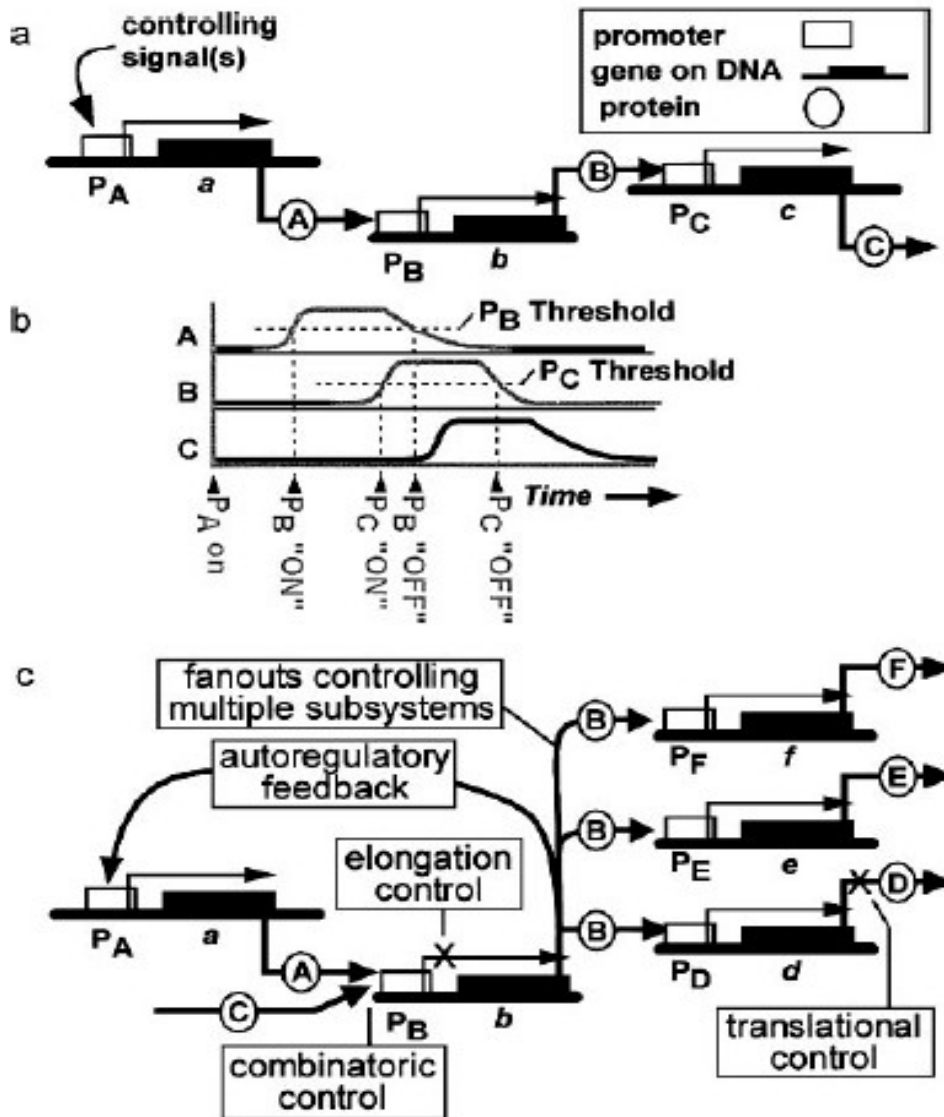
- cis-trans specificity
- small number of trans factors to a cis element: 8-10
- cis elements are programs
- regulation is event driven (asynchronous)
- regulation systems are noisy environments
- Protein-DNA and protein-protein regulation
- regulation changes with time

# **Gene Networks: models of measurable properties of Gene Regulatory Systems.**

Gene networks model functional elements of a Gene Regulation System together with the regulatory relationships among them in a computational formalism.

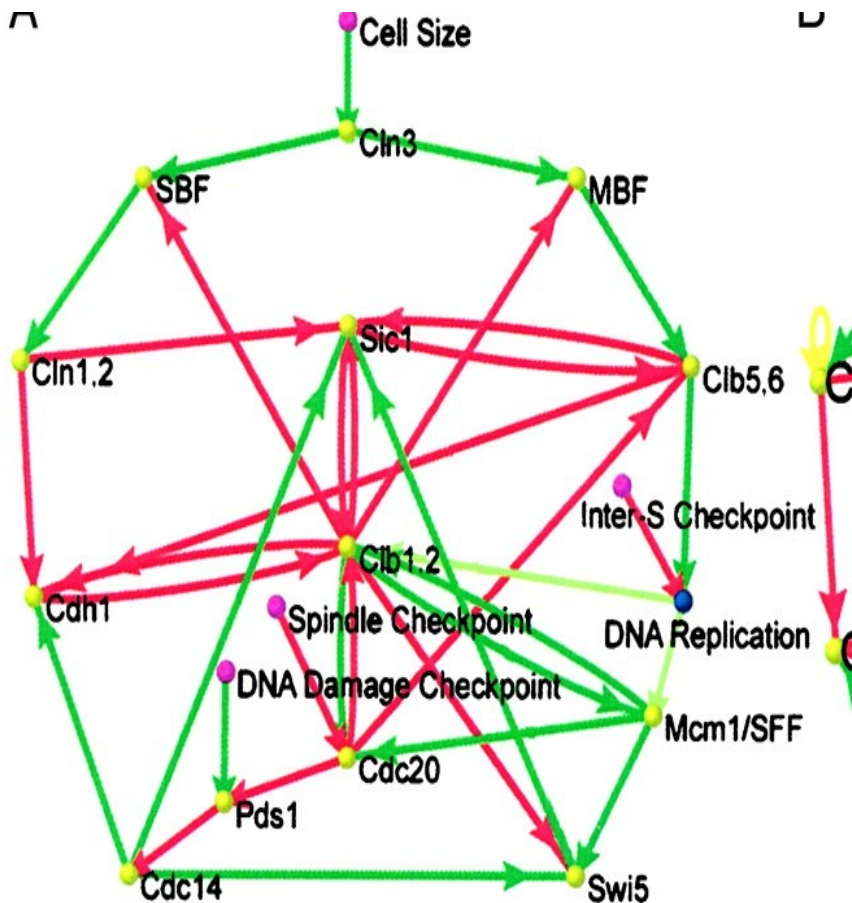
Types of relationships: causal, binding specificity, protein-DNA binding, protein-protein binding, etc.

# Simple Genetic Circuits



McAdams and Arkin et al 1998

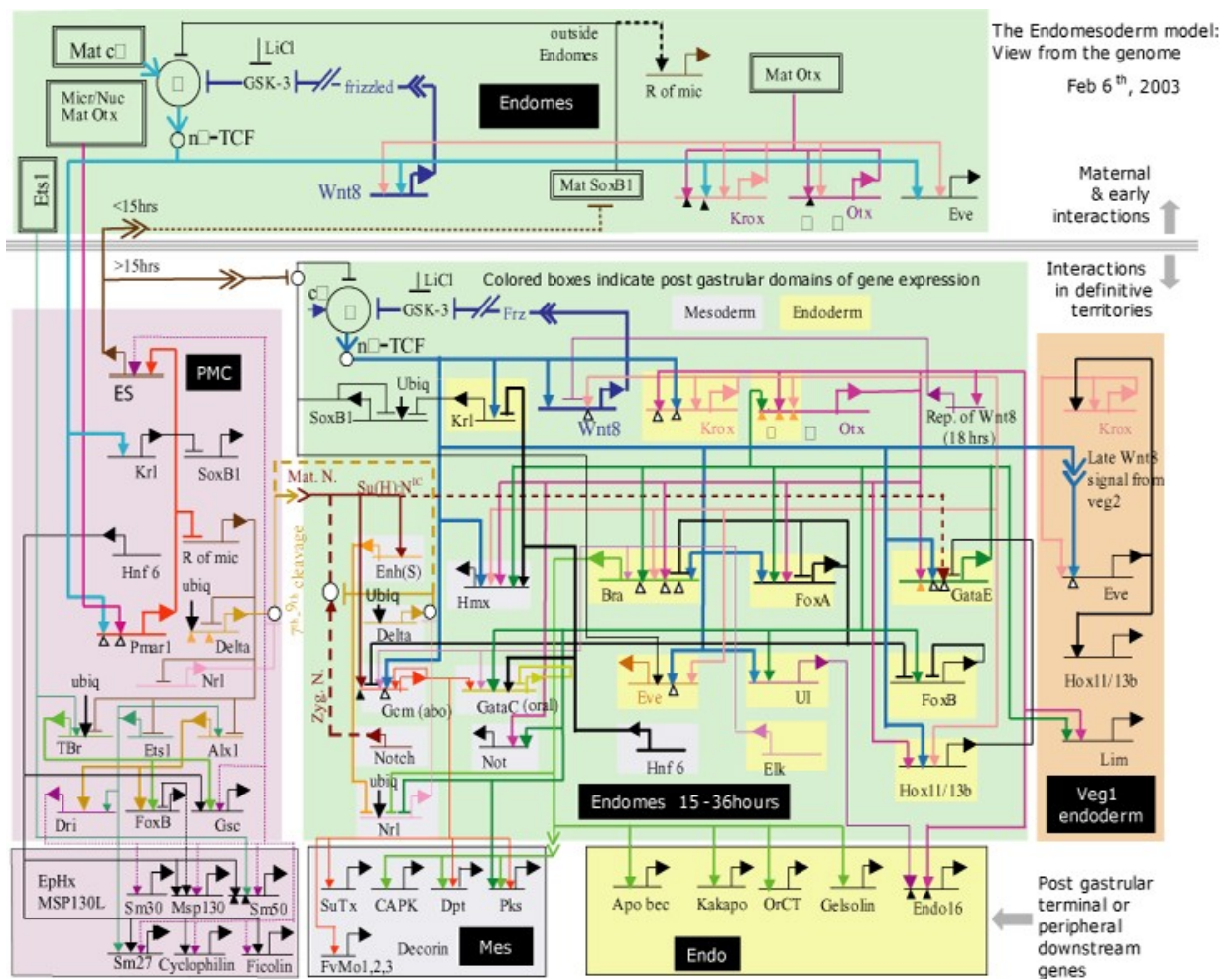
# Cell cycle network in *S. Cerevisiae*



Li, Fangting et al. (2004) Proc. Natl. Acad. Sci.

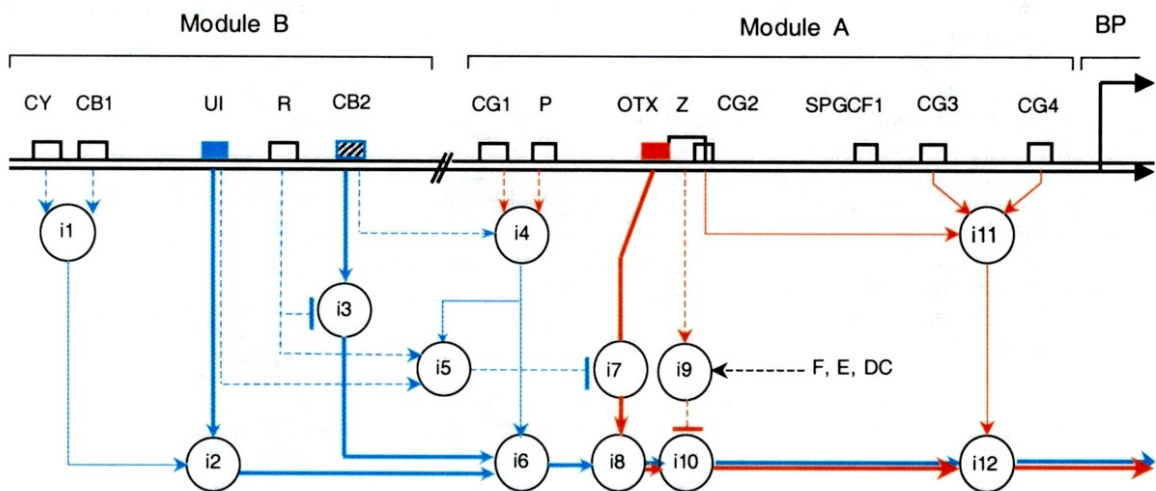


# Gene network of endomeso development in Sea Urchin



Davidson et al. Science 2002

# Logic of Cis-regulation



if CY & CB1 else	$i1 = 1$ $i1 = 0.5$	if $i5 = 0$ else	$i7 = OTX(t)$ $i7 = 0$
	$i2 = i1 \cdot UI(t)$		$i8 = i6 + i7$
if R else	$i3 = CB2(t)$ $i3 = k \cdot CB2(t)$ ( $1 < k < 2$ )	if (F or E or DC) & Z else	$i9 = 1$ $i9 = 0$
if P & CG1 & CB2 else	$i4 = 2$ $i4 = 0$	if $i9 = 1$ else	$i10 = 0$ $i10 = i8$
if $UI(t) > \text{threshold}$ & R & $i4 \neq 0$ else	$i5 = 1$ $i5 = 0$	if (CG2 & CG3 & CG4) else	$i11 = 2$ $i11 = 1$
	$i6 = i4 \cdot (i2 + i3)$		$i12 = i11 \cdot i10$



# Modeling Formalisms

## **Combinatorial (Qualitative)**

## **Physical (Quantitative or Continuous)**

- Static Graph Models
- Boolean Networks
- Weight Matrix (Linear) Models
- Bayesian Networks

- Stochastic Models
- Difference / Differential Equation Models
- Chemical/Physical Models
- Concurrency models

# Continuous Models of Gene Regulation

# Outline

- Quantitative Modeling
- Discrete vs. Continuous
- Modeling problems
- Models:
  - ODE
  - PDE
  - Stochastic
- Conclusions

# Quantitative Modeling in Biology

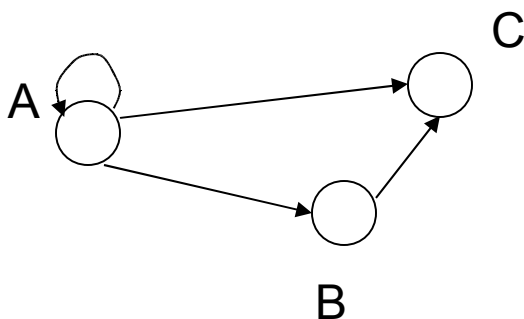
- State variables: concentrations of substances, e.g. proteins, mRNA, small molecules, etc.
- Knowing a system means being able to predict the concentrations of all key substances (state variables)
- Quantitative Modeling is the process of connecting the components of a system in a mathematical equation
- Solving the equations yields testable predictions for all state variables of the system

# Discrete vs. Continuous

- Here we will talk about continuous models, where values of variables change continuously in time (and/or space)
- On a molecular scale things are discrete, but on a macro scale they blend in and look continuous
- Next class we'll discuss discrete models

# Why Continuous?

- Continuous models are appealing because they allow for instantaneous change
- Continuous models let us express the precise relationships between instantaneous states of variables in a system



vs.

$$\frac{dA}{dt} = 1 - 2A$$

$$\frac{dB}{dt} = 0.5A$$

$$\frac{dC}{dt} = 2A + B$$

# Problems

When modeling with differential equations we face all the same problems as in the discrete models:

- Posing the equations. This presumes we understand the underlying phenomenon
- Data Fitting. How do we learn the model from the data?
- Solving the equations. Means we can do the math
- Model Behavior. Analyzing the fitted model to understand its behavior

# Recall the Modeling Process...

1. Knowledge
2. Modeling Objectives
3. Construct and Revise Models
4. Model behavior and predictions
5. Compare to new data
6. Better Models, goto 3
7. Learn...

# 1. Ordinary Differential Equations

Rate equation:

$$\frac{dx_i}{dt} = f_i(\mathbf{x}), \quad 1 \leq i \leq n$$

where

$\mathbf{x} = [x_1, \dots, x_n]$  is a vector of  $n$  concentrations

$f_i(x) : \mathbf{R}^n \rightarrow \mathbf{R}$  is a function

Systems of ODEs: There are  $n$  such equations

Solving the rate equations depends on  $f$ , but what is the form of the function  $f$ ?

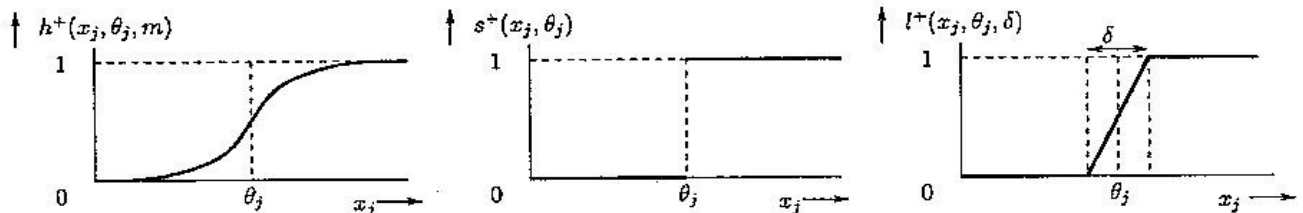
The answer is: as simple as possible.

# The Rate Function and Regulation

- The rate function specifies the interactions between the state variables.
- Its input are the concentrations, and the output is indicative (i.e. a function of) the change in a gene's regulation
- The regulation function describes how the concentration is related to regulation

$$h^+(x_j, \theta_j, m) = \frac{x_j^m}{x_j^m + \theta_j^m}, \quad (8)$$

- This is a typical regulation function, called a sigmoid, below compared to similar ones



# Non-linear ODEs

The rate function is nonlinear!

Eg.

1. Sigmoidal
2. Nonlinear, additive. Summarizes all pair wise (and nothing but pair wise) relationship

$$\frac{dX_i}{dt} = \sum_j T_{ij} f_j(X_j)$$

3. Nonlinear, non-additive. Summarizes all pairs and triplets of relationships

$$\frac{dX_i}{dt} = \sum_{jk} T_{ijk} f_j(X_j) f_k(X_k) + \sum_j T_{ij} f_j(X_j)$$

## Solving

- In general, these equations are difficult to solve analytically when  $f_i(\mathbf{x})$  are non-linear
- Numerical Simulators/Solvers work by numerically approximating the concentration values at discretized, consecutive time-points. Popular software for biochemical interactions:
  - **DBsolve**
  - **GEPASI**
  - **MIST**
  - **SCAMP**
- Although analytical solutions are impossible, we can learn a lot from general analyses of the behavior of the models, which some of the packages above provide

## Model Behavior:

- Feedback is essential in biological systems. The following is known about feedback:
  - negative feedback loops: system approach or oscillate around a single steady state
  - positive feedback loops: system tends to settle in one of two stable states
  - in general: a negative feedback loop is necessary for stable oscillation, and a positive feedback loop is necessary for multistationarity

## Data Fitting

- Fitting the parameters of a non-linear system is a difficult problem.
- Common solution: non-linear optimization scheme
  - explore the parameter space of the system
  - for each choice of parameters the models are solved numerically (e.g. Runge-Kutta)
  - the parameterized model is compared to the data with a goodness of fit function. It is this function that is optimized
- Genetic Algorithms and Simulated Annealing, with proper transition functions have been used with promising results

# Linear and Piecewise Linear ODEs

## Linear

- These are much easier to deal with: if the input variables are limited by a constant, they can be solved and learned polynomially, depending on the amount of data available

$$\frac{dX_i}{dt} = \sum_j w_{ij} X_j$$

- One way to learn them is by approximating them with linear weight models

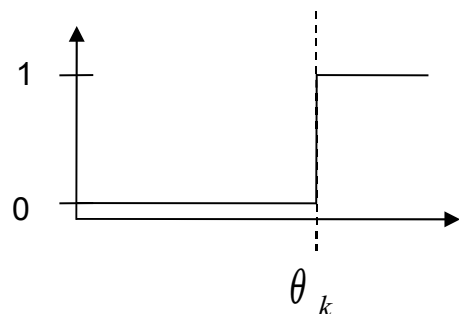
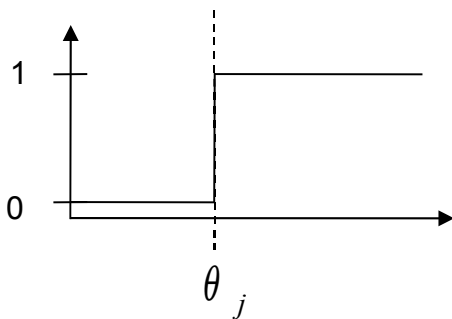
# Piecewise linear

- Approximating the sigmoid regulatory function with a step function

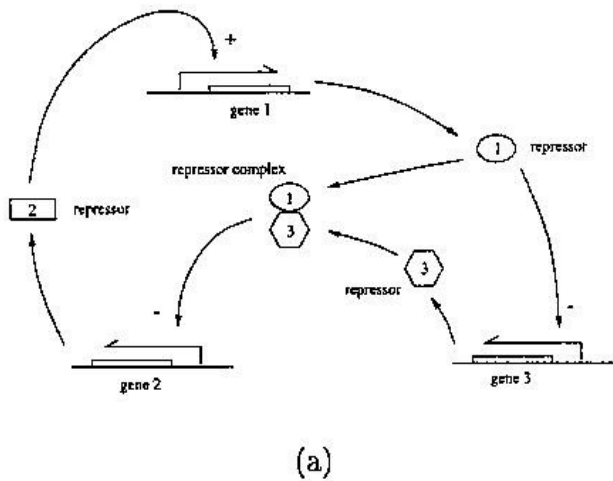
$$\frac{dX_i}{dt} = g_i(\mathbf{x}) - \gamma_i x_i, \quad 1 \leq i \leq n$$

$$g_i(\mathbf{x}) = \sum_{l \in L} k_{il} b_{il}(\mathbf{x}) \geq 0$$

- Here the function  $b_{il}$  is a function of  $n$  variables, defined in terms of sums and products of step functions:

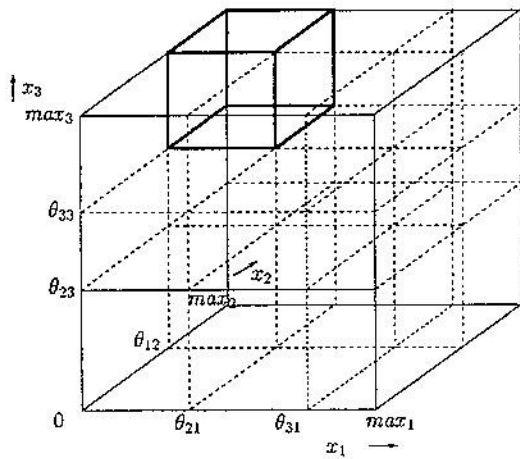


- This amounts to subdividing  $n$ -dimensional space into “orthants”, and in each of the orthants the PLODEs reduce to ODEs



$$\begin{aligned} \dot{x}_1 &= \kappa_1 s^+(x_2, \theta_{21}) - \gamma_1 x_1 \\ \dot{x}_2 &= \kappa_2 (1 - s^+(x_1, \theta_{11}) s^+(x_3, \theta_{31})) - \gamma_2 x_2 \\ \dot{x}_3 &= \kappa_3 s^-(x_1, \theta_{12}) + \kappa_4 s^-(x_3, \theta_{32}) - \gamma_3 x_3 \end{aligned} \quad (b)$$

**FIG. 9.** (a) Example regulatory network of three genes and (b) corresponding piecewise-linear differential equations;  $x_1$ ,  $x_2$ , and  $x_3$  represent protein or mRNA concentrations, respectively,  $\kappa_1, \dots, \kappa_4$  production constants,  $\gamma_1, \dots, \gamma_3$  degradation constants, and  $\theta_{11}, \theta_{12}, \theta_{21}, \theta_{31}, \theta_{32}$  threshold constants.



$$\begin{aligned} \dot{x}_1 &= \kappa_{12} - \gamma_1 x_1 \\ \dot{x}_2 &= -\gamma_2 x_2 \\ \dot{x}_3 &= \kappa_{31} - \gamma_3 x_3 \end{aligned} \quad (b)$$

**FIG. 10.** (a) The phase space box of the model in Fig. 9, divided into  $2 \cdot 3 \cdot 3 = 18$  orthants by the threshold planes. (b) The state equations for the orthant  $0 \leq x_1 < \theta_{21}$ ,  $\theta_{12} < x_2 \leq \max_2$ , and  $\theta_{33} < x_3 \leq \max_3$  (the orthant demarcated by bold lines).

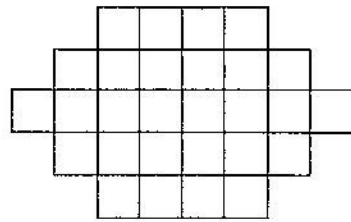
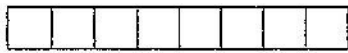
de Jong, JCB 2002

## 2. PDES

- ODEs count on spatial homogeneity
- In other words, ODEs don't care where the processes take place
- But in some real situation this assumption clearly does not hold
  - Diffusion
  - Transcription factor gradients in development
  - Multicellular organisms

# Example: Reaction-Diffusion Equations

$$\frac{dx_i^{(l)}}{dt} = f_i(\mathbf{x}^{(l)}) + \delta_i (x_i^{(l+1)} - 2x_i^{(l)} + x_i^{(l-1)}), \quad 1 \leq i \leq n, \quad 1 < l < p. \quad (16)$$



The equation above describes the change in conc. for all state variables, in all cells of the line above. When the number of cells is large, this becomes a PDE:

$$\frac{\partial x_i}{\partial t} = f_i(\mathbf{x}) + \delta_i \frac{\partial^2 x_i}{\partial l^2}, \quad 0 \leq l \leq \lambda, \quad 1 \leq i \leq n. \quad (17)$$

If it is assumed that no diffusion occurs across the boundaries  $l = 0$  and  $l = \lambda$ , the boundary conditions become

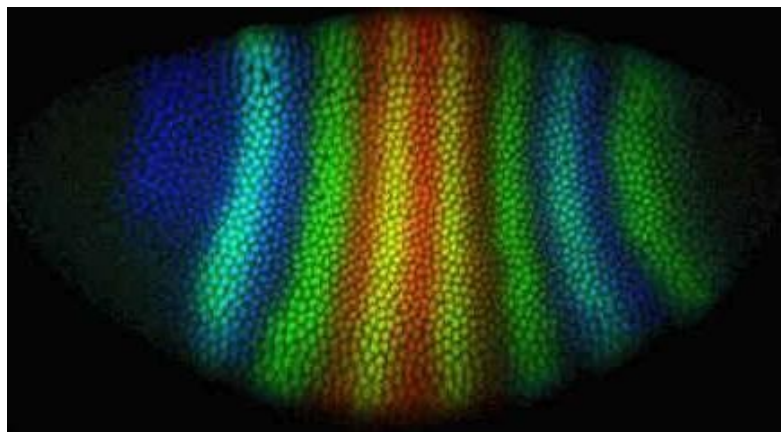
$$\frac{\partial^2}{\partial l^2} x_i(0, t) = 0 \quad \text{and} \quad \frac{\partial^2}{\partial l^2} x_i(\lambda, t) = 0. \quad (18)$$

These equations were first introduced in the study of developmental phenomena and pattern formation by Turing.

Direct analytical solutions are impossible even for two variables ( $n=2$ )

# Drosophila Example

- These PDE models have been used repeatedly to model developmental examples in the fruit fly
- Instances of the reaction-diffusion equations (only more specific) have been used to model the striped patterns in a drosophila embryo



# 3. Stochastic Master Equations

- Deterministic modeling is not always possible, but also sometimes incorrect
- Assumptions of deterministic, continuous models:
  - Concentrations of substances vary deterministically
  - Conc. of substances vary continuously
- On molecular level, both assumptions may not be correct
- Solution: Instead of deterministic values, accept a joint probability distribution, similar to the one discussed in the Bayesian Network lectures.

# Equation:

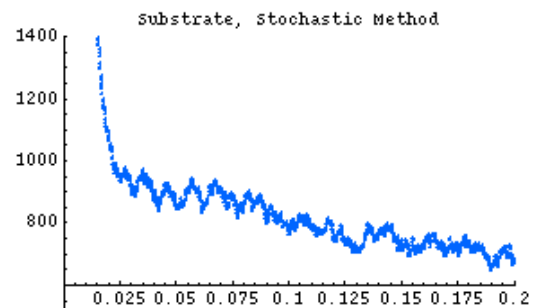
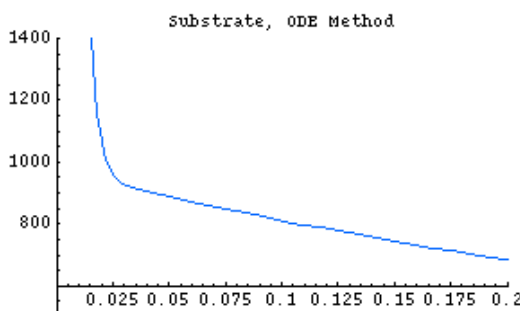
species, etc. The time evolution of the function  $p(\mathbf{X}, t)$  can now be specified as follows:

$$p(\mathbf{X}, t + \Delta t) = p(\mathbf{X}, t) \left( 1 - \sum_{j=1}^m \alpha_j \Delta t \right) + \sum_{j=1}^m \beta_j \Delta t, \quad (21)$$

where  $m$  is the number of reactions that can occur in the system,  $\alpha_j \Delta t$  the probability that reaction  $j$  will occur in the interval  $[t, t + \Delta t]$  given that the system is in the state  $\mathbf{X}$  at  $t$ , and  $\beta_k \Delta t$  the probability that reaction  $j$  will bring the system in state  $\mathbf{X}$  from another state in  $[t, t + \Delta t]$  (Gillespie, 1977, 1992). Rearranging (21), and taking the limit as  $\Delta t \rightarrow 0$ , gives the *master equation* (van Kampen, 1997):

$$\frac{\partial}{\partial t} p(\mathbf{X}, t) = \sum_{j=1}^m (\beta_j - \alpha_j p(\mathbf{X}, t)). \quad (22)$$

These equations are very difficult to solve and simulate!



(c) Jason Kastner and Caltech

ODE vs. Stochastic solutions

# Biography

- Hidde de Jong, Modeling and Simulation of Genetic Regulatory Systems: A Literature Review. *Journal of Computational Biology* 9(1): 67-103 (2002).
- McAdams and Arkin, *Annu. Rev. Biophys. Biomol. Struct.* 1998 (27)
- Setty, Y. et al., Detailed map of a cis-regulatory input function, *PNAS*, 100:7702-7707 (2003)
- Fangting et al., *PNAS* 2004 (101)
- Albert, R. and Othmer, H.G. *Journal of Theoretical Biology* 223, 1-18 (2003).
- Davidson, E.H. et al., *Science* 295, 1669-1678, 2002
- D. C. Weaver and C. T. Workman and G. D. Stromo, *Pacific Symposium on Biocomputing*, 1999.
- D'Haeseleer et al., *Pacific Symposium on Biocomputing*, 1999.