Sequence Analysis

- ECS129
- Patrice Koehl

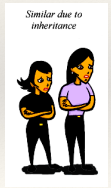
Sequence Analysis: Outline

1. Why do we compare sequences?
2. Sequence comparison: from qualitative to quantitative methods
3. Deterministic methods: Dynamic programming
4. Heuristic methods: BLAST
5. Multiple Sequence Alignment

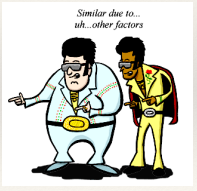
Similarity: Homology vs Analogy

Homology: **Similarity in characteristics resulting from shared ancestry.**

Analogy: **The similarity of characteristics between two species that are not closely related; attributable to convergent evolution.**



Two sisters: homologs



Two "Elvis": analogs

Homology: Orthologs and Paralogs

Homology: Similarity in characteristics resulting from shared ancestry.

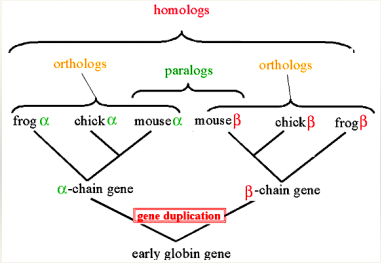
Paralogy: Homologous sequences are paralogous if they were separated by a gene duplication event

Orthology: Homologous sequences are orthologous if they were separated by a speciation event

Further reading:

Koonin EV (2005). "Orthologs, paralogs, and evolutionary genomics". Annu. Rev. Genet. 39:309-338.

Homology: Orthologs and Paralogs



Applications of Sequence Analysis

- Sequencing projects, assembly of sequence data
- Evolutionary history
- Identification of functional elements in sequences
- gene prediction
- Classification of proteins
- Comparative genomics
- RNA structure prediction
- Protein structure prediction
- Health Informatics

DNA sequence: Chargaff's rules

Rule 1: In double stranded DNA, the amount of guanine is equal to cytosine and the amount of adenine is equal to thymine

(basis of Watson Crick base pairing)

Rule 2: the composition of DNA varies from one species to another; in particular in the relative amounts of A, G, T, and C bases

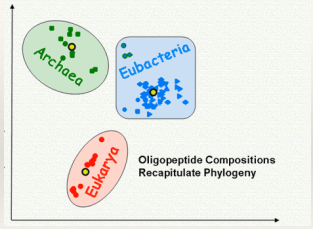
DNA sequence: Chargaff's rules

Table 3-2 Data Leading to the Formulation of Chargaff's Rules

Source	Adenine to Guanine	Thymine to Cytosine	Adenine to Thymine	Guanine to Cytosine	Purines to Pyrimidines
Ox	1.29	1.43	1.04	1.00	1.1
Human	1.56	1.75	1.00	1.00	1.0
Hen	1.45	1.29	1.06	0.91	0.99
Salmon	1.43	1.43	1.02	1.02	1.02
Wheat	1.22	1.18	1.00	0.97	0.99
Yeast	1.67	1.92	1.03	1.20	1.0
Hemophilus influenzae	1.74	1.54	1.07	0.91	1.0
E-coli K2	1.05	0.95	1.09	0.99	1.0
Avian tubercle bacillus	0.4	0.4	1.09	1.08	1.1
Serratia marcescens	0.7	0.7	0.95	0.86	0.9
Bacillus schatz	0.7	0.6	1.12	0.89	1.0

SOURCE: After E. Chargaff et al., *J. Biol. Chem.* 177 (1949).

Comparing sequences based on their tri-peptide content



Proteins: Structure, Function and Genetics **54**, 20-40 (2004)

Comparing individual letters

Scores are usually stored in a "weight" matrix also called "substitution" matrix or "matching" matrix.

Defining the "proper" matrix is still an active area of research:

1. Identity matrix
2. Chemical property matrix
   In this matrix amino acids or nucleotides are intuitively classified on the basis of their chemical properties
3. Substitution-based matrix
   Dayhoff matrix
   PAM matrices
   Blosum matrices

Substitution Matrices

Dayhoff matrix was created in 1978 based on few closely related (> 85% identity) sequences available this time (1500 aligned amino-acids).

PAM-family of matrices is a simple update of the original Dayhoff matrix.

Gonnet matrices were created by exhaustive alignment of all Database sequences in 1992.

BLOSUM matrix is based on local similarities (blocks) of proteins rather than overall alignments.

Most common Scoring Matrices

BLOSUM matrices ***(Henikoff and Henikoff, 1992)***

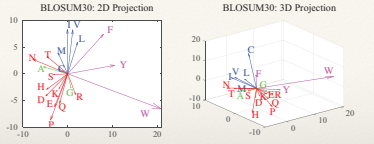
- Start from "reliable" alignments of sequences with at least XX % identity
- Compute mutation probabilities
- Convert into Scores: -> BLOSUMXX matrix

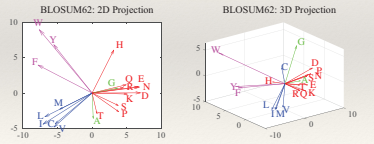
PAM matrices ***(Dayhoff, 1974)***

- Point Accepted Mutation
- Start with PAM score = 1: alignments of sequences with 1 mutation -> PAM1 matrix
- Generate successive PAM matrices: PAMXX = $(PAM1)^{XX}$

Example of a Scoring matrix: Blosum62

	C	S	T	P	A	G	N	D	E	Q	H	R	K	M	I	L	V	F	Y	W
C	9	-1	-1	-3	0	-3	-3	-3	-4	-3	-3	-3	-3	-1	-1	-1	-1	-2	-2	-2
S	-1	4	1	-1	1	0	1	0	0	0	-1	-1	0	-1	-2	-2	-2	-2	-2	-3
T	-1	1	4	1	-1	1	0	1	0	0	0	-1	0	-1	-2	-2	-2	-2	-2	-3
P	-3	-1	1	7	-1	-2	-1	-1	-1	-1	-2	-2	-1	-2	-3	-3	-2	-4	-3	-4
A	0	1	-1	-1	4	0	-1	-2	-1	-1	-2	-1	-1	-1	-1	-1	-1	-2	-2	-3
G	-3	0	1	-2	0	6	-2	-1	-2	-2	-2	-2	-2	-3	-4	-4	0	-3	-3	-2
N	-3	1	0	-2	-2	0	6	1	0	0	-1	0	0	-2	-3	-3	-3	-3	-2	-4
D	-3	0	1	-1	-2	-1	1	6	2	0	-1	-2	-1	-3	-3	-4	-3	-3	-3	-4
E	-4	0	0	-1	-1	-2	0	2	5	2	0	0	1	-2	-3	-3	-3	-3	-2	-3
Q	-3	0	0	-1	-1	-2	0	0	2	5	0	1	1	0	-3	-2	-2	-3	-1	-2
H	-3	-1	0	-2	-2	-2	1	1	0	0	8	0	-1	-2	-3	-3	-2	-1	2	-2
R	-3	-1	-1	-2	-1	-2	0	-2	0	1	0	5	2	-1	-3	-2	-3	-3	-2	-3
K	-3	0	0	-1	-1	-2	0	-1	1	1	-1	2	5	-1	-3	-2	-3	-3	-2	-3
M	-1	-1	-1	-2	-1	-3	-2	-3	-2	0	-2	-1	-1	5	1	2	-2	0	-1	-1
I	-1	-2	-2	-3	-1	-4	-3	-3	-3	-3	-3	-3	-3	1	4	2	1	0	-1	-3
L	-1	-2	-2	-3	-1	-4	-3	-4	-3	-2	-3	-2	-2	2	2	4	3	0	-1	-2
V	-1	-2	-2	-2	0	-3	-3	-3	-2	-2	-3	-3	-2	1	3	1	4	-1	-1	-3
F	-2	-2	-2	-4	-2	-3	-3	-3	-3	-3	-1	-3	-3	0	0	0	-1	6	3	1
Y	-2	-2	-2	-3	-2	-3	-2	-3	-2	-1	2	-2	-2	-1	-1	-1	-1	3	7	2
W	-2	-3	-3	-4	-3	-2	-4	-4	-3	-2	-2	-3	-3	-1	-3	-2	-3	1	2	11





DotPlot: Overview of Sequence Similarity

Build a table S:

- rows: Sequence 1
- columns: Sequence 2

Assign a score S(i,j) to each entry in the table:

- select a window size WS

- Compare window around i with window around j -> Score(i,j)

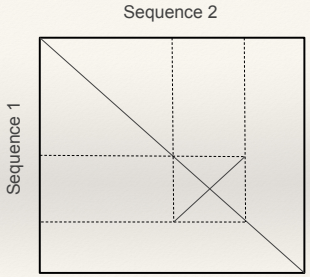
Display table of scores S

- show a dot at position (i,j) if Score(i,j) > Threshold

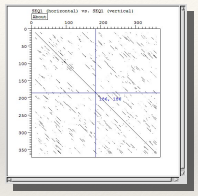
Patterns on DotPlot

Internal Repeat Insertion (Deletion) Divergence

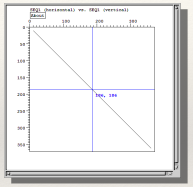
Patterns on DotPlot



Patterns on DotPlot



With many details



Overall view - no details

What is sequence alignment?

Given two sequences of letters and a scoring scheme for evaluating letter matching, find the optimal pairing of letters from one sequence to the other.

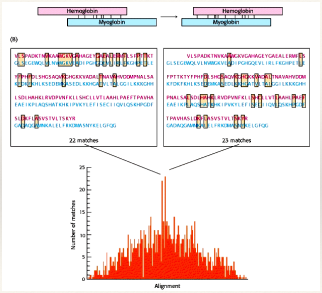
Human hemoglobin (α chain)

VLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHG
SAQVKGHGKKVADALTNAVAHVDDMPNALSALSDLHAHKLRVDPVNFKLLS
HCLLVTLAAHLPAEFTPAVHASLDKFLASVSTVLTSKYR

Human myoglobin

GLSDGEWQLVLNVWGKVEADIPGHGQEVLIRLFKGHPETLEKFDKFKHLKS
EDEMKASEDLKKHGATVLTALGGILKKKGHHEAEIKPLAQSHATKHKIPVK
YLEFISECIIQVLQSKHPGDFGADAQGAMNKALELFRKDMASNYKELGFQG

Ungapped Alignment



(From Biochemistry, Stryer, fifth edition)

Alignment with gap(s)

How do we generate the "best" gapped alignment ?

Total number of possible gapped alignment: $\sum_{k=1}^{\min(N,M)} \binom{N}{k}\binom{M}{k}$

DP and Sequence Alignment

Key idea:

The score of the optimal alignment that ends at a given pair of positions in the sequences is the score of the best alignment previous to these positions plus the score of aligning these two positions.

DP and Sequence Alignment

Test all alignments that can lead to i aligned with j

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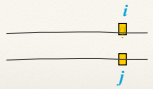
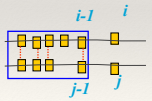
Test all alignments that can lead to i aligned with j



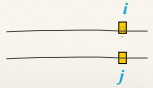
3 possibilities:

1) i-1 aligned with j-1



DP and Sequence Alignment

Test all alignments that can lead to i aligned with j



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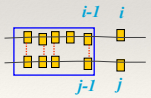
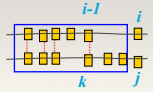
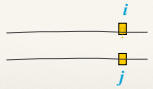


2) i-1 aligned with k, 1≤k ≤j-2



DP and Sequence Alignment

Test all alignments that can lead to i aligned with j



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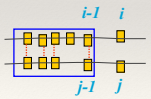

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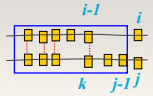
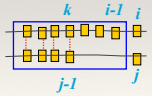


3) j-1 aligned with l, 1≤l ≤i-2



DP and Sequence Alignment

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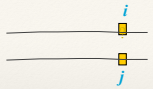

3 possibilities:

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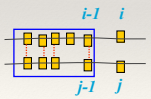
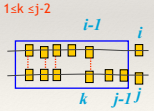


2) i-1 aligned with k, 1≤k ≤j-2



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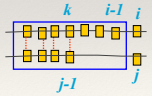


Choose option that leads to the best score

Implementing the DP algorithm for sequences

Aligning 2 sequence S1 and S2 of lengths N and M:

1) Build a NxM alignment matrix A such that A(i,j) is the optimal score for alignments up to the pair (i,j)
2) Find the best score in A
3) Track back through the matrix to get the optimal alignment of S1 and S2.

Implementing the DP algorithm for sequences

Aligning 2 sequence S1 and S2 of lengths N and M:

Example

Sequence 1: AWVCDEC

Sequence 2: AWEC
Score(i,j) = 10 if i=j, 0 otherwise

no gap penalty

Example

1) Initialize

	A	W	V	C	D	E	C
A	10	0	0	0	0	0	0
W	0						
E	0						
C	0						

Example

2) Propagate

	A	W	V	C	D	E	C
A	10	0	0	0	0	0	0
W	0	20					
E	0						
C	0						

Example

2) Propagate

	A	W	V	C	D	E	C
A	10	0	0	0	0	0	0
W	0	20	10				
E	0						
C	0						

Example

2) Propagate

	A	W	V	C	D	E	C
A	10	0	0	0	0	0	0
W	0	20	10	10	10	10	10
E	0	10	20	20	20	30	20
C	0	10	20	30	20		

Example

3) Trace back

	A	W	V	C	D	E	C
A	10	0	0	0	0	0	0
W	0	20	10	10	10	10	10
E	0	10	20	20	20	30	20
C	0	10	20	30	20	20	40

Alignment:

AWVCDEC

AW------EC

Total score: 40

Example 2

	A	A	T	G	C
A	10	10	0	0	0
G	0	10	10	20	10
G	0	10	10	20	20
C	0	10	10	10	30

Alignments:

High Score: 30

AATGC AATGC AATGC AATG C AATG C

AG GC A GGC AGGC A GGC A GGC

Example 3

Gap cost: -2

		A	A	T	G	C
A	10	8	-2	-2	-2
G	-2	10	8	18	8
G	-2	8	10	18	16
C	-2	8	8	10	28

Alignments:

High Score: 28

AATGC AATGC AATGC

AG GC A GGC AGGC

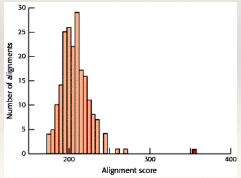
Statistical Significance of alignment: Shuffling

Score: 355

Shuffling a sequence:

THISISTHECORRECTSEQUENCE

TSTCRTQNHIHOESUCISERCEEE



Gap penalty

Most common model:

$$W_N = G_0 + N * G_1$$

W_N: gap penalty for a gap of size N

G_0 : cost of opening a gap

G_1 : cost of extending the gap by one

N : size of the gap

Global versus Local Alignment

Global alignment finds the arrangement that maximizes total score

Best known algorithm: Needleman and Wunsch.

Local alignment identifies highest scoring subsequences,
sometimes at the expense of the overall score.

Best known algorithm: Smith and Waterman.

Local alignment algorithm is just a variation of the global alignment algorithm!

Modifications for local alignment

1) The scoring matrix has negative values for mismatches

1) The minimum score for any (i,j) in the alignment matrix is 0.

1) The best score is found anywhere in the filled alignment matrix

These 3 modifications cause the algorithm to search for matching sub-sequences which are not penalized by other regions (modif. 2), with minimal poor matches (modif 1), which can occur anywhere (modif 3).

Global versus Local Alignment

Match: +1; Mismatch: -2; Gap: -1

	A	C	C	T	G	S
A	1	-3	-3	-3	-3	-3
C	-3	2	1	-2	-2	-2
C	-3	1	3	-1	-1	-1
N	-3	-2	-1	1	0	0
S	-3	-2	-1	0	-1	1

	A	C	C	T	G	S
A	1	0	0	0	0	0
C	0	2	1	0	0	0
C	0	1	3	0	0	0
N	0	0	0	1	0	0
S	0	0	0	0	0	1

Global: **ACCTGS** **ACCTGS**
ACC-NS **ACCN-S**

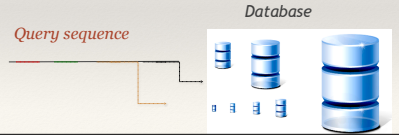
Local: **ACC**
ACC

BLAST

(Basic Local Alignment Search Tool)

Main ideas:

1. Construct a list of all words in the query sequence
2. Scan database for sequences that contain one or more of the query words
3. Initiate a local alignment for each word match between query and database



Original BLAST

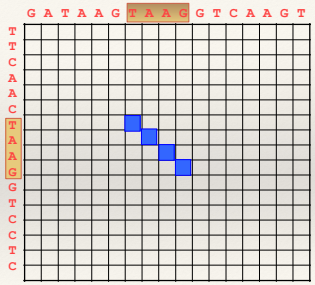
1. Define dictionary
   All words of length k
   (typically k=11)
2. Scan database sequences for matches
   with alignment score ≥ T
   (typically T = k)
3. Generate alignment
   ungapped extensions until score
   below statistical threshold
4. Output all local alignments with scores
   above the statistical threshold

Original BLAST

An example:

k = 4, T = 4

1) The matching word TAAG initiates an alignment



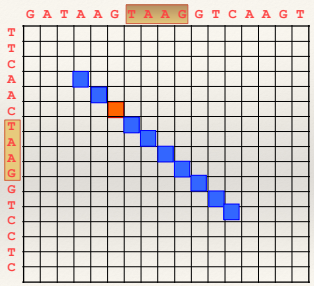
Original BLAST

An example:

k = 4, T = 4

1) The matching word AGGT initiates an alignment

2) Extension of the alignment to the left and right with no gap until alignment score falls below 50%



Original BLAST

An example:

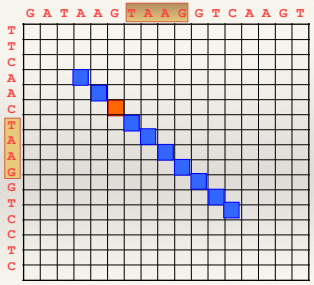
k = 4, T = 4

1)The matching word AGGT initiates an alignment

2)Extension of the alignment to the left and right with no gap until alignment score falls below 50%

3)Output:

AAGTAAGGTC
AACTAAGGTC



Gapped BLAST

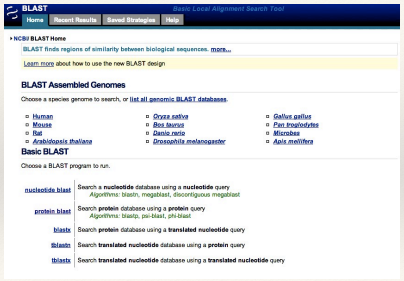
An example:

k = 4, T = 4

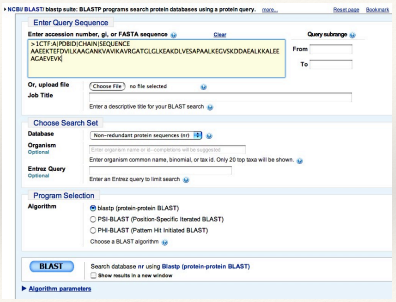
1)The matching word GGTC initiates an alignment

2) Extend alignment in a band around anchor

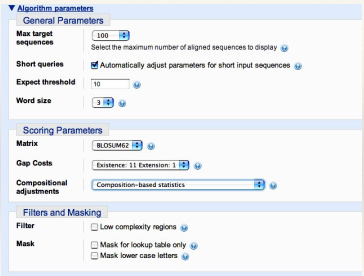
BLAST Portal



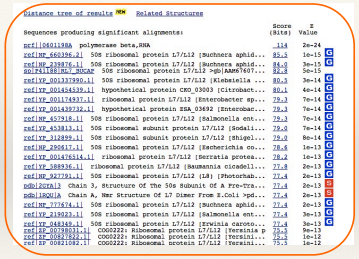
BLAST: Input



BLAST Parameters



BLAST Results



Distance tree of results NEW Related Structures

Sequences producing significant alignments:

	Score (Bits)	E Value
pdb|1G61|19A polymerase beta,RNA	114	2e-24
ref|NP_660396.2| 50S ribosomal protein L7/L12 [Buchnera aphid...	85.5	1e-15
ref|NP_239876.1| 50S ribosomal protein L7/L12 [Buchnera aphid...	84.0	3e-15
sp|P41188|RL7_BUCAP 50S ribosomal protein L7/L12 >gb|AAM67407...	82.8	5e-15
ref|YP_001337990.1| 50S ribosomal protein L7/L12 [Klebsiella ...	80.5	3e-14
ref|YP_001454339.1| hypothetical protein CKO_03003 [Citrobact...	80.1	4e-14
ref|YP_001174937.1| ribosomal protein L7/L12 [Enterobacter sp...	79.3	7e-14
ref|YP_001439732.1| hypothetical protein ESA_03692 [Enterobac...	79.3	7e-14
ref|NP_457918.1| 50S ribosomal protein L7/L12 [Salmonella ent...	79.3	7e-14
ref|YP_453813.1| 50S ribosomal subunit protein L7/L12 [Sodali...	79.0	7e-14
ref|YP_312899.1| 50S ribosomal subunit protein L7/L12 [Shigel...	79.0	8e-14
ref|NP_290617.1| 50S ribosomal protein L7/L12 [Escherichia co...	78.6	1e-13
ref|YP_001476514.1| ribosomal protein L7/L12 [Serratia protea...	78.2	1e-13
ref|YP_588936.1| ribosomal protein L7/L12 [Baumannia cicadell...	77.8	2e-13
ref|NP_927791.1| 50S ribosomal protein L7/L12 (L8) [Photorhab...	77.4	2e-13
pdb|2GYA|3 Chain 3, Structure Of The 50s Subunit Of A Pre-Tra...	77.4	2e-13
pdb|1RQU|A Chain A, Nmr Structure Of L7 Dimer From E.Coli >pd...	77.4	2e-13
ref|NP_777474.1| 50S ribosomal protein L7/L12 [Buchnera aphid...	77.4	2e-13
ref|YP_219023.1| 50S ribosomal protein L7/L12 [Salmonella ent...	77.4	3e-13
ref|YP_048342.1| 50S ribosomal protein L7/L12 [Erwinia caroto...	77.4	3e-13
ref|ZP_00788361.1| COG0222: Ribosomal protein L7/L12 [Yersinia p	75.5	9e-13
ref|ZP_00821082.1| COG0222: Ribosomal protein L7/L12 [Yersini...	75.5	1e-12
ref|YP_00821082.1| COG0222: Ribosomal protein L7/L12 [Yersini...	75.5	1e-12

Statistics of Protein Sequence Alignment

- ***Statistics of global alignment:***

  Unfortunately, not much is known! Statistics based on Monte Carlo simulations (shuffle one sequence and recompute alignment to get a distribution of scores)

- ***Statistics of local alignment***

  Well understood for ungapped alignment. Same theory probably apply to gapped-alignment

Statistics of Protein Sequence Alignment

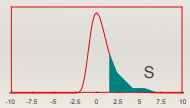
What is a local alignment ?

"Pair of equal length segments, one from each sequence,
whose scores can not be improved by extension or trimming.
These are called high-scoring pairs, or HSP"

http://www.people.virginia.edu/~wrp/cshl98/Altschul/Altschul-1.html

The E-value for a sequence alignment

HSP scores follow an extreme value distribution, characterized by two parameters, K and λ.



The expected number of HSP with score at least S is given by:

$$E = Kmn\exp(-\lambda S)$$

m, n : sequence lengths

E : E-value

The Bit Score of a sequence alignment

Raw scores have little meaning without knowledge of the scoring scheme used for the alignment, or equivalently of the parameters K and λ.

Scores can be normalized according to:

$$S' = \frac{\lambda S - \ln(K)}{\ln(2)}$$

S' is the **bit score** of the alignment.

The E-value can be expressed as:

$$E = mn2^{-S'}$$

The P-value of a sequence alignment

The number of random HSP with score greater of equal to S follows a Poisson distribution:

$$P(X \text{ random HSP with score} \geq S) = \exp(-E)\frac{E^X}{X!}$$

(E: E-value)

Then:

$$P(0 \text{ random HSP with score} \geq S) = \exp(-E)$$

$$P_{val} = P(\text{at least } 1 \text{ random HSP with score} \geq S) = 1 - \exp(-E)$$

Note: when E <<1, P ≈E

The database E-value for a sequence alignment

2) Longer sequences are more likely to be related to the query:

$$E_{DB} = N_S K m n \exp(-\lambda S)$$

BLAST reports E_{DB2}

$$E_{DB2} = K m N_R \exp(-\lambda S)$$

Why multiple sequence alignment?

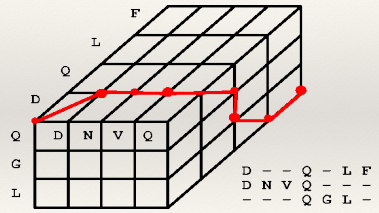
```
Seq1: AALGCLVKDYFPEP--VTVSWNSG---
Seq2: VSLTCLVKGFYPSD--IAVEWWSNG--
```

Why multiple sequence alignment?

```
Seq1: AALGCLVKDYFPEP--VTVSWNSG---
Seq2: VSLTCLVKGFYPSD--IAVEWWSNG--
Seq3: VTISCTGSSSNIGAG-NHVKWYQQLPG
Seq4: VTISCTGTSSNIGS--ITVNWYQQLPG
Seq5: LRLSCSSSGFIFSS--YAMYWVRQAPG
Seq6: LSLTCTVSGTSFDD--YYSTWVRQPPG
Seq7: PEVTCVVVDVSHEDPQVKFNWYVDG--
Seq8: ATLVCLISDFYPGA--VTVAWKADS--
```

MSA: Dynamic programming?

Theoretically, it is possible to extend the dynamic programming technique to N sequences.




```
D – – Q – L F
D N V Q – – –
– – – Q G L –
```

MSA: Dynamic programming?

- One of the most important properties of an algorithm is how its execution time increases as the problem is made larger.
This is **the computational complexity** of the algorithm

- There is a notation to describe the algorithmic complexity, called **the big-O notation**.
If we have a problem of size (i.e. number of input data points) n, then an algorithm takes **O(n)** time if the time increases linearly with n.

-It is important to realize that an algorithm that is **quick on small problems may be totally useless on large problems** if it has a bad O() behavior.

MSA: Dynamic programming?

Standard description of algorithms, where n is the size of the problem, and c is a constant:

Complexity	Type	Computing time for n=1000 (1 operation=1s)
O(c)	Dream…	Seconds
O(log(n))	Really good	10 seconds
O(n)	good	1000 seconds = 5 mins
$O(n^2)$	Not so good	10^6 seconds = 11.5 days
$O(n^3)$	Bad	10^9 seconds = 31 years
$O(c^n)$	Catastrophic!	Millions of years!!

MSA: Dynamic programming?

Computational complexity of dynamic programming:

-Two sequences of length M : **$O(M^2)$**

-Three sequences of length M: **$O(M^3)$**

- N sequences of length M: **$O(M^N)$**

-> dynamic programming is not a reasonable option for aligning multiple sequences!

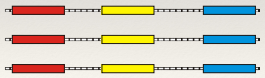
MSA: Approximate methods

1. **Progressive global alignment**
   Start with the most similar sequences and builds the alignment by adding the rest of the sequences
2. **Iterative methods**
   Start by making alignments of small group of sequences and then revise the alignment for better results
3. **Alignment based on small conserved domains**
4. **Alignment based on statistical or probabilistic models of the sequence**

Multiple sequence alignment: using conserved domains

Sequences often contain highly conserved regions

These regions can be used for an initial alignment



How to generate a multiple sequence alignment?

Raw Alignment

Human	N Y L S
Chimp	N K Y L S
Gorilla	N F S
Orangutan	N F L S

How to generate a multiple sequence alignment?

Sequence elements are not truly independent but related by phylogeny:

Raw Alignment

Human	N Y L S
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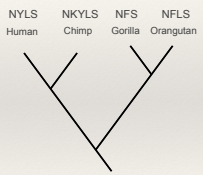

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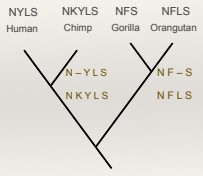

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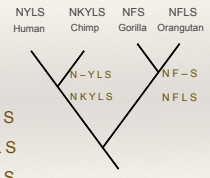

N – Y L S
N K Y L S
N – F – S
N – F L S

Multiple sequence alignment: Progressive method

A) Perform pairwise alignments

	HAHU	HBHU	HAHO	HBHO	MYWHP	PILHB	LOHB
HAHU							
HBHU	21.1						
HAHO	32.9	19.7					
HBHO	20.7	**39.0**	20.4				
MYWHP	11.0	9.8	10.3	9.7			
PILHB	9.3	8.6	9.6	8.4	7.0		
LOHB	7.1	7.3	7.5	7.4	7.3	4.3	

Multiple sequence alignment: Progressive method

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	HAHU	HBHU	HAHO	HBHO	MYWHP	PILHB	LOHB
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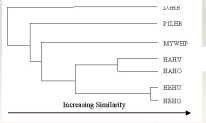
PILHB	9.3	8.6	9.6	8.4	7.0		
LOHB	7.1	7.3	7.5	7.4	7.3	4.3	

B) Cluster based on similarity



Multiple sequence alignment: Progressive method

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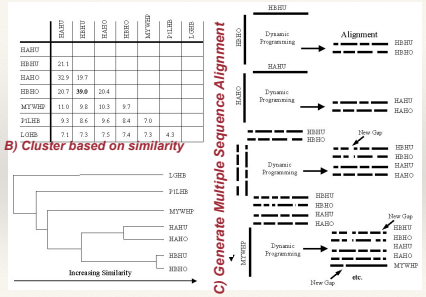

	HAHU	HBHU	HAHO	HBHO	MYWHP	PALHB	LGHB
HAHU							
HBHU	21.1						
HAHO	32.9	19.7					
HBHO	20.7	**39.0**	20.4				
MYWHP	11.0	9.8	10.3	9.7			
PALHB	9.3	8.6	9.6	8.4	7.0		
LGHB	7.1	7.3	7.5	7.4	7.3	4.3	

B) Cluster based on similarity

C) Generate Multiple Sequence Alignment

Some References on Alignments

Global Alignment:

Needleman, S.B. and Wunsch, C.D. (1970). "A general method applicable to the search for similarities in the amino acid sequence of two proteins". *Journal of Molecular Biology* ***48 (3): 443–53***

Local alignment:

Smith, T.F. and Waterman, M.S. (1981) "Identification of Common Molecular Subsequences". *Journal of Molecular Biology* **147: 195–197**

ClustalW:

Thompson, J. D., Higgins, D.G. and Gibson, T.J. (1994) "CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, positions-specific gap penalties and weight matrix choice". *Nucleic Acids Research*, **22:4673-4680**

What have we learnt?

1) **Sequence analysis** is one of the keys that will help us unravel the information coming from Genomics

2) **Vocabulary**

Analogy: The similarity of characteristics between two species that are not closely related

Homology: Similarity in characteristics resulting from shared ancestry

- **Paralog:** Homologous sequences are paralogous if they were separated by a gene duplication event
- **Ortholog:** Homologous sequences are orthologous if they were separated by a speciation event

3) In bioinformatics we often assume that **sequence similarity implies homology**. However we do need to be cautious.

What have we learnt?

4) Sequence analysis starts with **an analysis of its content**

1) DNAs:

Chargaff rule2: the composition of DNA varies from one species to another

2) Proteins:

Tri-peptide content identifies the kingdom of life (bacteria, archea or eukaryot)

5) **DotPlots** are very useful, qualitative tools for sequence comparison

4) **Scoring** between sequences is usually based on **substitution matrices**

Most common matrices: PAM and BLOSUM

What have we learnt?

1. **Dynamic programming (DP)** is an algorithm for aligning two sequences that is guaranteed to generate the optimal alignment, under the hypothesis that the scores are additive.
2. There are two variants of DP used for sequence analysis
   Global alignment: Needleman and Wunsch
   Local alignment: Smith and Waterman
3. DP is too slow for comparing a sequence with a large database
4. **BLAST** provides a heuristic method for detecting sequences that are similar
5. **BLAST is best for detection** and should not be trusted for the alignment itself

What have we learnt?

6) Multiple sequence alignment: definition

A multiple sequence alignment is an alignment of n > 2 sequences obtained by inserting gaps ("-") into sequences such that the resulting sequences have all length L. MSW can help to reveal biological facts about proteins, to establish homology,…

7) Difficulties in generating MSA

Most pairwise alignment algorithms are too complex to be used for N-wise alignments

8) Three main types of MSA algorithms:

- Progressive global alignment (starts with the most alike sequences)
  * e.g., ClustalW, ClustalX
- Iterative methods (initial alignment of groups of sequences that are revised)
  * MultAlin, PRRP, SAGA
- Alignments based on locally conserved patterns